**INVITED REVIEW** 

# Nitric oxide synthases in the pathogenesis of cardiovascular disease

Lessons from genetically modified mice

Hiroaki Shimokawa • Masato Tsutsui

Received: 7 January 2010/Revised: 27 January 2010/Accepted: 28 January 2010/Published online: 24 February 2010 © Springer-Verlag 2010

Abstract Nitric oxide (NO) is produced in almost all tissues and organs, exerting a variety of biological actions under both physiological and pathological conditions. NO is synthesized by three distinct NO synthase (NOS) isoforms (neuronal, inducible, and endothelial NOS), all of which are expressed in the human cardiovascular system. Although the regulatory roles of NOSs in cardiovascular diseases have been described in pharmacological studies with selective and non-selective NOS inhibitors, the specificity of the NOS inhibitors continues to be an issue of debate. To overcome this issue, genetically engineered animals have been used. All types of NOS gene-deficient animals, including singly, doubly, and triply NOS-deficient mice, and various types of NOS gene-transgenic (TG) animals, including conditional and non-conditional TG mice bearing endothelium-specific or cardiomyocytespecific overexpression of each NOS gene, have thus been developed. The roles of individual NOS isoforms as well as the entire NOS system in the cardiovascular system have been extensively investigated in those mice, providing pivotal insights into an understanding of the pathophysiology of NOSs in human cardiovascular diseases. Based on studies with the murine NOS genetic models, this review briefly summarizes the latest knowledge of NOSs and cardiovascular diseases.

H. Shimokawa (⊠) Department of Cardiovascular Medicine, Tohoku University Graduate School of Medicine, 1-1Seiryo-machi, Aoba-ku, Sendai 980-8574, Japan e-mail: shimo@cardio.med.tohoku.ac.jp

M. Tsutsui Department of Pharmacology, Ryukyu University, Okinawa, Japan Keywords Cardiovascular diseases · Knockout mice · Nitric oxide · Nitric oxide synthase · Transgenic mice · Endothelium · Endothelium-derived relaxing factor (EDRF) · NO

## Introduction

Nitric oxide (NO) plays an important role in maintaining cardiovascular homeostasis through multiple biological actions [12, 21, 48, 65, 76, 78]. NO is formed from its precursor L-arginine by a family of NO synthases (NOSs) with stoichiometric production of L-citrulline. The NOS system consists of three distinct isoforms, including neuronal (nNOS or NOS-1), inducible (iNOS or NOS-2), and endothelial NOS (eNOS or NOS-3).

It was initially demonstrated that nNOS and eNOS are constitutively expressed mainly in the nervous system and the vascular endothelium, respectively, synthesizing a small and physiological amount of NO in a calcium-dependent manner both under basal conditions and upon stimulation, whereas iNOS is induced only when stimulated by microbial endotoxins or certain proinflammatory cytokines, producing a greater amount of NO in a calcium-independent manner [12, 21, 48, 65, 76, 78]. However, it was subsequently demonstrated that nNOS and eNOS are also subject to expressional regulation [9, 11, 51, 52, 73] whereas iNOS also could be constitutively expressed even under physiological conditions [4, 57].

Genetically engineered animals are a powerful experimental tool to study the function of target genes in vivo. All types of NOS gene-deficient animals, including singly, doubly, and triply NOS-deficient mice, have been developed (Table 1) [13, 14, 18, 19, 35, 38, 44, 56, 64, 68, 72, 83]. Furthermore, various types of NOS gene-transgenic (TG) animals, including conditional and non-conditional

Pflugers Arch - Eur J Physiol (2010) 459:959-967

Table 1 Mice lacking the NOS genes that have thus far been established   (Modified from Ref. [76] with a permission)	NOS <sup>-/-</sup> mice	Sites of gene deletion	References
	nNOS <sup>-/-</sup>	Exon 2 (#1)	[18]
		Exon 6	[14]
		Exon 6	[56]
	iNOS <sup>-/-</sup>	Proximal 585 bases of promoter plus exons 1-4 (#2)	[38]
		Near exons 1–5	[83]
		Exons 12 and 13 and a part of exon 11 (#3)	[35]
	eNOS <sup>-/-</sup>	Exons 24–26 (#4)	[19]
		Exon 12 (#5)	[64]
		Exons 24 and 25	[13]
	n/iNOSs <sup>-/-</sup>	#1 and #3	[72]
		#1 and #2	[44]
	n/eNOSs <sup>-/-</sup>	#1 and #4	[68]
		#1 and #5	[72]
		#1 and #4	[44]
<i>NOS</i> nitric oxide synthase, <i>nNOS</i> neuronal NOS, <i>iNOS</i> inducible NOS, <i>eNOS</i> endothelial NOS	i/eNOSs <sup>-/-</sup>	#3 and #5	[72]
		#2 and #4	[44]
	n/i/eNOSs <sup>-/-</sup>	#1, #2 and #4	[44]

TG mice with endothelium-specific or cardiomyocytespecific overexpression of each NOS isoform, have also been established (Table 2) [3, 5, 16, 22, 37, 46, 53, 55, 70, 77]. By using those genetically modified mice, the cardiovascular roles of NOSs have been extensively studied, and the findings provide important insights into the significance of NOSs in human cardiovascular diseases. In this review, we briefly summarize the current knowledge of NOSs and cardiovascular diseases on the findings obtained from the NOS gene-modified mice.

# Vascular lesion formation

# Role of eNOS

Endothelium-specific eNOS-TG mice with an 8-fold increase in vascular NOS activity showed decreased neointimal formation after carotid artery ligation [25] and another strain of endothelium-specific eNOS-TG mice with a 10-fold increase in vascular NOS activity similarly exhibited a reduction in atherosclerotic vascular lesion formation induced by breeding with  $ApoE^{-/-}$  mice [77]. Consistent with these findings, eNOS<sup>-/-</sup> mice showed increased neointimal formation, accelerated medial thickening, and abnormal vascular remodeling in response to permanent carotid artery ligation (Fig. 1) [45, 85] and cuff placement around the femoral artery [59]. Furthermore, eNOS<sup>-/-/</sup>ApoE<sup>-/-</sup> mice had accelerated formation of atherosclerotic vascular lesions as compared with ApoE<sup>-/-</sup> mice [26, 30]. These lines of evidence indicate vasculoprotective roles of eNOS in the pathogenesis of atherosclerotic vascular lesion formation. In contrast, in endotheliumspecific eNOS-TG mice with an 8-fold increase in vascular NOS activity, atherosclerotic vascular lesion formation is accelerated when crossbred with  $ApoE^{-/-}$  mice, where

Table 2 Mice overexpressing   the NOS gene that have thus NOS-TG mice   far been established nNOS-TG   iNOS-TG iNOS-TG	NOS-TG mice	Overexpression site	Promoter used	References
	nNOS-TG	Myocardium(conditional)	α-MHC	[5]
	Myocardium(conditional)	α-MHC	[37]	
		Brain	CaMKIIα	[55]
	iNOS-TG	Myocardium(conditional)	α-MHC	[46]
		Myocardium	α-MHC	[16]
(Modified from Ref. [76] with a permission)		Pancreatic $\beta$ cell	Insulin	[70]
	eNOS-TG	Endothelium	Preproendothelin-1	[53]
TG transgenic. MHC myosin	nsgenic. MHC myosin	Endothelium	eNOS	[77]
heavy chain, <i>CaMKII</i> calcium- calmodulin multifunctional ki- nase II		Myocardium	α-MHC	[3]
		Myocardium	α-MHC	[22]



Fig. 1 The different vasculoprotective roles of three nitric oxide synthase (NOS) isoforms in a mouse carotid artery ligation model. Studies with each NOS isoform-deficient mice demonstrated that endothelial NOS (eNOS) inhibits neointimal formation, that inducible NOS (iNOS) attenuates constrictive vascular remodeling, and that neuronal NOS (nNOS) suppresses both neointimal formation and constrictive vascular remodeling. Thus, individual NOS isoforms have different vasculoprotective actions against vascular lesion formation in mice in vivo.–inhibition. (Modified from Ref. [76] with a permission)

enhanced oxidative stress due to relative tetrahydrobiopterin deficiency and development of a tolerance of the vascular smooth muscle to NO were demonstrated [54].

In eNOS<sup>-/-</sup> mice, in addition to the abolishment of NO-mediated relaxations, endothelium-dependent responses (relaxations and hyperpolarizations) mediated by endothelium-derived hyperpolarizing factor (EDHF) are markedly reduced [40]. Furthermore, the EDHF-mediated responses also are sensitive to catalase [40] and also are markedly reduced in Cu,Zn-SOD<sup>-/-</sup> mice [42]. These lines of evidence indicate that endothelium-derived hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), which is formed through dismutation of eNOS-derived superoxide anions by Cu,Zn-SOD, is an EDHF [40, 42, 66], although it is highly possible that several different factors and mechanisms other than endothelium-derived H<sub>2</sub>O<sub>2</sub> derived from NOSs are also involved in the EDHF-mediated responses.

## Role of iNOS

The role of iNOS in vascular lesion formation seems to be complicated. Deletion of the iNOS gene in mice exacerbated pathological vascular remodeling in a carotid artery ligation model (Fig. 1) [85] and in a cardiac transplant model [27]. However, the iNOS deletion conversely ameliorated neointimal formation in a carotid cuff placement model [7] and lipid-rich atherosclerotic vascular lesion formation in ApoE<sup>-/-</sup> mice [29]. Thus, iNOS appears to have two faces, which could be explained in part by the

oxidant and antioxidant properties of iNOS. Indeed, NOSs produce superoxide anions rather than NO, with a resultant production of a potent oxidant peroxynitrite, under certain pathological conditions such as deficiency of a substrate (e.g., L-arginine) or a cofactor (e.g., tetrahydrobiopterin) termed as pathological NOS uncoupling [80, 81].

## Role of nNOS

Although the regulatory roles of eNOS and iNOS in vascular lesion formation have been widely studied, little has been known about the role of nNOS. However, the expression of nNOS also is up-regulated in the neointima, endothelial cells, and macrophages in both early and advanced human atherosclerotic lesions [84]. We demonstrated that in nNOS<sup>-/-</sup> mice neointimal formation and constrictive vascular remodeling (a reduction in vascular cross-sectional area) following carotid artery ligation are accelerated (Fig. 1) [43]. In agreement with our finding, nNOS<sup>-/-/</sup>ApoE<sup>-/-</sup> mice showed accelerated atherosclerotic vascular lesion formation as compared with ApoE<sup>-/-</sup> mice [31]. Up-regulation of nNOS may play a compensatory role in the presence of reduced eNOS activity (e.g., inflammation and arteriosclerosis) to maintain vascular homeostasis [73]. Furthermore, we demonstrated that inflammatory and proliferative stimuli and statins increase vascular nNOS expression [51, 52]. It also has been reported that hypoxic conditions [82] and hypertension [2] up-regulate vascular nNOS expression.

## Role of the whole NOSs system

Because all NOSs play a role in the vascular system, we next conceived a project to investigate the roles of the whole NOS system in vivo. The roles of the NOS system in the human body have been investigated in pharmacological studies with non-selective NOS inhibitors and in studies with NOS isoform-deficient mice. However, because of both the non-specificity of the NOS inhibitors and compensation among the three NOS isoforms, the authentic roles of the NOS system were still poorly understood. To address this important issue, we have recently developed mice in which the entire NOS system is completely disrupted (triply nNOS/iNOS/eNOS-deficient mice) [44, 50]. The triply n/i/eNOS<sup>-/-</sup> mice, but not any singly NOS<sup>-/-</sup> mice, spontaneously develop arteriosclerotic vascular lesions (neointimal formation, medial thickening, and perivascular fibrosis) in the coronary and renal arteries, and lipid-rich atherosclerotic vascular lesions in the aorta, even on a normal chow diet [50, 74]. These results provided the first direct evidence for a vasculoprotective role of the entire NOS system in atherosclerosis.

## Spontaneous myocardial infarction

Myocardial infarction (MI) is the leading cause of death for both genders worldwide [1, 28]. However, the molecular mechanisms for the pathogenesis of MI remain to be fully elucidated.

## Role of the whole NOSs system

Although eNOS has potent vasculoprotective effects, [12, 21, 48, 65, 76, 78] neither deletion of the eNOS gene nor pharmacological inhibition of eNOS activity induces MI in animals. In contrast, our triply n/i/eNOSs<sup>-/-</sup> mice spontaneously develop MI and sudden cardiac death (Fig. 2a,b) [50, 74, 75]. This is the first in vivo demonstration of the protective roles of NOS synthases in the pathogenesis of MI. In our triply n/i/eNOSs<sup>-/-</sup> mice, arteriosclerosis is noted in most of the vasculature,

whereas atherosclerosis is observed in the aorta alone. In humans, MI is caused not only by coronary atherosclerosis, but also by other mechanisms, including coronary intimal inflammation and coronary vasospasm [1, 79]. In the triply n/i/eNOSs<sup>-/-</sup> mice that died of MI, marked coronary intimal hyperplasia and medial thickening were noted (Fig. 2b,c). Furthermore, in the dead triply n/i/ eNOSs<sup>-/-</sup> mice, marked infiltration of mast cells at the coronary artery adventitia was observed (Fig. 2d). Histamine released from adventitial mast cells is thought to cause coronary vasospasm with a resultant MI in humans [32]. It is thus possible that coronary arteriosclerosis and coronary vasospasm are involved in the cause of MI and death in the triply NOSs<sup>-/-</sup> mice (Fig. 3) [50, 74]. In our triply n/i/eNOSs-/- mice, not only NO-mediated but also EDHF-mediated endothelium-dependent relaxations are abolished [69] in addition the enhanced contractions to phenylephrine [74]. These vascular dysfunctions may also



Fig. 2 Decreased survival, spontaneous myocardial infarction (MI), coronary arteriosclerosis and mast cell infiltration in male triply  $n/i/eNOSs^{-/-}$  mice. **a** Survival rate (n=29-57). A red line represents markedly reduced survival in the triply  $n/i/eNOSs^{-/-}$  mice. \*, †, and #P<0.05 between wild-type (WT) C57BL/6J vs. singly, doubly, and triply NOS-KO, respectively. **b** Acute MI and coronary arteriosclerotic lesion formation in the triply  $n/i/eNOSs^{-/-}$  mouse that died at 8 months of age (Masson-trichrome staining). *Blue* in the heart cross-section of

the dead triply n/i/eNOSs<sup>-/-</sup> mouse indicates antero-septal acute MI. Adjacent coronary artery shows marked luminal narrowing, wall thickening, and perivascular fibrosis (*blue*). **c** Arteriosclerotic lesion formation in serial sections of the infarct-related coronary artery. **d** Mast cell infiltration in the coronary artery adventitia (toluidine-blue staining) n=10-33. *Red arrows* indicate mast cells. \**P*<0.05 vs. WT. (Reproduced from Ref. [76] with a permission)



Fig. 3 Mechanisms for spontaneous MI caused by the defective NOS system in mice in vivo. Genetic disruption of all NOSs causes metabolic syndrome, hypoadiponectinemia, hyper-low-density-lipoprotein (LDL)emia, coronary adventitial mast cell infiltration, and vascular dysfunction. These factors could contribute to the pathogenesis of spontaneous MI. Importantly, long-term pharmacological blockade of the AT<sub>1</sub> receptor significantly reduces the incidence of MI, along with amelioration of these risk factors. Thus, it is possible that the AT<sub>1</sub> receptor pathway is substantially involved in the molecular mechanisms of the pathological phenotypes and MI in the triply  $n/i/eNOSs^{-/-}$  mice. (Modified from Ref. [76] with a permission)

be involved in the pathogenesis of MI in the triply  $NOSs^{-/-}$  mice (Fig. 3).

#### Metabolic syndrome

Metabolic syndrome (MetS) is defined as a pathological state with accumulated cardiovascular risk factors of metabolic origin, including visceral obesity, hypertension, dyslipidemia, impaired glucose tolerance, and insulin resistance [49, 71]. Importantly, accumulation of three or more risk factors dramatically increases the risk of morbidity of atherosclerotic cardiovascular diseases by 11-fold, indicating that MetS is an important therapeutic target for the prevention and treatment of cardiovascular diseases [49, 71].

Roles of eNOS and the whole NOSs system

eNOS<sup>-/-</sup> and our triply n/i/eNOSs<sup>-/-</sup> mice manifest phenotypes that closely resemble MetS in humans [50, 74, 76]. Although the extent of each of cardiovascular risk factors (hypertension, dyslipidemia, and visceral obesity) was comparable between the 2 genotypes, the extent of impaired glucose tolerance and that of insulin resistance were greater in the triply n/i/eNOSs<sup>-/-</sup> than in the eNOS<sup>-/-</sup> mice, and hyper-low-density-lipoproteinemia was observed only in the triply n/i/eNOSs<sup>-/-</sup> mice [50]. Thus, it is possible that the whole NOSs system plays important roles in the prevention of MetS [50]. Although metabolic risk factors are present in the two genotypes, spontaneous MI is noted only in the triply  $n/i/eNOSs^{-/-}$  mice. This discrepancy may be related to a compensatory mechanism by other NOSs that are not genetically disrupted [68]. Indeed, in the  $eNOS^{-/-}$  mice, up-regulation of vascular nNOS expression has been reported [17, 33]. Furthermore, we also have demonstrated that NOS activity and plasma NOx levels are fairly well preserved in the singly  $eNOS^{-/-}$  mice [44].

Adiponectin is an anti-atherogenic adipocytokine, which improves dyslipidemia, glucose metabolism, and insulin resistance, and inhibits the progression of atherosclerosis [24, 41, 67]. Under the condition of obesity with adipocyte hypertrophy, the synthesis of adiponectin is decreased and in patients with MetS, the circulating levels of adiponectin are also reduced, in contrast to the increases in other atherogenic adipocytokine levels. This adiponectin deficiency is thought to play a pivotal role in the pathogenesis of MetS and its vascular complications [41]. In our triply n/i/eNOSs<sup>-/-</sup> mice, plasma adiponectin levels were significantly reduced [50]. Thus, the adiponectin deficiency may contribute to the development of metabolic abnormalities and arteriosclerotic lesion formation in the triply n/i/eNOS<sup>-/-</sup> mice (Fig. 3).

Importantly, the renin-angiotensin system is markedly activated in the triply  $n/i/eNOSs^{-/-}$  mice, and long-term treatment with an angiotensin II type 1 (AT1) receptor blocker, olmesartan, potently inhibit coronary arteriosclerotic lesion formation, adventitial mast cell infiltration, and the occurrence of MI in the mice, with a resultant improvement in prognosis [50]. Furthermore, the long-term treatment with olmesartan reverses all the abnormal metabolic phenotypes, together with amelioration of hypo-adiponectinemia [50]. These results suggest that the AT1 receptor pathway is substantially involved in the pathogenesis of MI in our triply  $n/i/eNOSs^{-/-}$  mice (Fig. 3).

## Heart failure

# Role of eNOS

In cardiomyocyte-specific eNOS-TG mice with a 30-fold increase in cardiac NOS activity, left ventricular (LV) remodeling after coronary artery ligation is suppressed, showing improved LV systolic and diastolic function and attenuation of LV hypertrophy [22]. Endothelium-specific eNOS-TG mice with a 12-fold increase in vascular NOS activity also exhibit improved survival, LV dysfunction, and pulmonary edema following coronary ligation without affecting LV remodeling [23]. Consistent with these findings,  $eNOS^{-/-}$  mice with heart failure (HF) due to either MI [63] or pressure overload [20] show reduced survival and exacerbation of LV remodeling and LV dysfunction. It also has been reported that eNOS mediates, at least in part, the beneficial cardiovascular protective effects of statins [34], angiotensin converting enzyme inhibitors [36], AT1 receptor blockers [36], and corticosteroids [15] in experimental HF. Thus, it is evident that eNOS plays important protective roles in HF [39, 58].

## Role of nNOS

Conditionally targeted cardiomyocyte-specific nNOS-TG mice with a 5-fold increase in cardiac NOS activity showed delayed transition toward HF in response to pressure overload [37]. In agreement with this evidence, two strains of  $nNOS^{-/-}$  mice with MI-induced HF similarly showed reduced survival and exacerbation of pathological LV remodeling or dysfunction after coronary artery ligation, although not totally identical in the two strains [8, 62]. Thus, it is possible that in addition to eNOS, nNOS also plays a cardioprotective role in HF [6].

## Role of iNOS

Increased iNOS expression is noted in cardiomyocytes in septic shock, myocarditis, ischemia, and dilated cardiomyopathy, and has been implicated in the development of HF. However, cardiomyocyte-specific iNOS overexpression per se (in two different strains with either a 10-fold [46] or 40-fold increase [16] in cardiac NOS activity) did not cause HF, suggesting that increased iNOS expression per se is not the triggering mechanism in HF. In contrast,  $iNOS^{-/-}$  mice with HF induced by MI [10, 60] and by pressure overload [86] showed improved survival, less LV remodeling and dysfunction, and decreased myocardial apoptosis. Furthermore, iNOS<sup>-/-</sup> mice with HF induced by cardio-specific overexpression of TNF- $\alpha$  show improved  $\beta$ -adrenergic inotropic responsiveness. Thus, it is possible that, in contrast to eNOS and nNOS, iNOS exerts opposite and unfavorable effects in HF. The underlying mechanisms for the contrasting roles of NOS isoforms in HF are unclear, but may relate to the differences in their spatial localization, expressional regulation, NO-generating capacity, and peroxynitrite generation [47, 58, 61].

# Other forms of cardiovascular diseases

A lines of accumulating evidence also has suggested that the impairment of the NOSs system is involved in the pathogenesis of other forms of cardiovascular diseases, including aortic aneurysms, arrhythmias, and congenital heart disease [76].

## Conclusions

The mouse is the most ideal genetically modifiable mammalian presently available. Studies with mice that are deficient of or overexpressing NOSs provide pivotal insights into the roles of NOSs in the pathogenesis of cardiovascular diseases. In general, eNOS and nNOS exert cardiovascular protective roles, while iNOS seems to exert dual effects in the cardiovascular system. The observations with the genetically modified animals have greatly advanced our understanding of the roles of the NOSs system in the pathogenesis of human cardiovascular diseases. Further studies are certainly needed to clarify whether these observations can be translated to human patients with cardiovascular diseases.

Acknowledgments This work was supported in part by the Grantin-Aid for Scientific Research on Innovative Areas (Signaling Functions of Reactive Oxygen Species), the Grant-in-Aid for Tohoku University Global COE for Conquest of Signal Transduction Diseases with Network Medicine, and the Grants-in-Aid for Scientific Research, all of which are from the Ministry of Education, Culture, Sports, Science and Technology, Tokyo, Japan.

#### References

- 1. Antman EM, Braunwald E (2005) ST-elevation myocardial infarction: pathology, pathophysiology, and clinical features. In: Zipes DP, Libby P, Bonow RO, Braunwald E (eds) Braunwald's heart disease, 7th edn. Elsevier Saunders, Philadelphia, pp 1141–1166
- Boulanger CM, Heymes C, Benessiano J, Geske RS, Levy BI, Vanhoutte PM (1998) Neuronal nitric oxide synthase is expressed in rat vascular smooth muscle cells: activation by angiotensin II in hypertension. Circ Res 83:1271–1278
- Brunner F, Andrew P, Wolkart G, Zechner R, Mayer B (2001) Myocardial contractile function and heart rate in mice with myocyte-specific overexpression of endothelial nitric oxide synthase. Circulation 104:3097–3102
- Buchwalow IB, Podzuweit T, Bocker W, Samoilova VE, Thomas S, Wellner M et al (2002) Vascular smooth muscle and nitric oxide synthase. FASEB J 16:500–508
- Burkard N, Rokita AG, Kaufmann SG, Hallhuber M, Wu R, Hu K et al (2007) Conditional neuronal nitric oxide synthase overexpression impairs myocardial contractility. Circ Res 100: e32–e44
- Casadei B (2006) The emerging role of neuronal nitric oxide synthase in the regulation of myocardial function. Exp Physiol 91:943–955
- Chyu KY, Dimayuga P, Zhu J, Nilsson J, Kaul S, Shah PK et al (1999) Decreased neointimal thickening after arterial wall injury in inducible nitric oxide synthase knockout mice. Circ Res 85:1192–1198
- Dawson D, Lygate CA, Zhang MH, Hulbert K, Neubauer S, Casadei B (2005) nNOS gene deletion exacerbates pathological left ventricular remodeling and functional deterioration after myocardial infarction. Circulation 112:3729–3737
- Dudzinski DM, Igarashi J, Greif D, Michel T (2006) The regulation and pharmacology of endothelial nitric oxide synthase. Annu Rev Pharmacol Toxicol 46:235–276

- Feng Q, Lu X, Jones DL, Shen J, Arnold JM (2001) Increased inducible nitric oxide synthase expression contributes to myocardial dysfunction and higher mortality after myocardial infarction in mice. Circulation 104:700–704
- Forstermann U, Boissel JP, Kleinert H (1998) Expressional control of the constitutive isoforms of nitric oxide synthase (NOS I and NOS III). FASEB J 12:773–790
- Furchgott RF (1984) The role of endothelium in the responses of vascular smooth muscle to drugs. Annu Rev Pharmacol Toxicol 24:175–197
- Godecke A, Decking UK, Ding Z, Hirchenhain J, Bidmon HJ, Godecke S et al (1998) Coronary hemodynamics in endothelial NO synthase knockout mice. Circ Res 82:186–194
- Gyurko R, Leupen S, Huang PL (2002) Deletion of exon 6 of the neuronal nitric oxide synthase gene in mice results in hypogonadism and infertility. Endocrinology 143:2767–2774
- 15. Hafezi-Moghadam A, Simoncini T, Yang Z, Limbourg FP, Plumier JC, Rebsamen MC et al (2002) Acute cardiovascular protective effects of corticosteroids are mediated by nontranscriptional activation of endothelial nitric oxide synthase. Nat Med 8:473–479
- Heger J, Godecke A, Flogel U, Merx MW, Molojavyi A, Kuhn-Velten WN et al (2002) Cardiac-specific overexpression of inducible nitric oxide synthase does not result in severe cardiac dysfunction. Circ Res 90:93–99
- Huang A, Sun D, Shesely EG, Levee EM, Koller A, Kaley G (2002) Neuronal NOS-dependent dilation to flow in coronary arteries of male eNOS-KO mice. Am J Physiol 282:H429–H436
- Huang PL, Dawson TM, Bredt DS, Snyder SH, Fishman MC (1993) Targeted disruption of the neuronal nitric oxide synthase gene. Cell 75:1273–1286
- Huang PL, Huang Z, Mashimo H, Bloch KD, Moskowitz MA, Bevan JA et al (1995) Hypertension in mice lacking the gene for endothelial nitric oxide synthase. Nature 377:239–242
- 20. Ichinose F, Bloch KD, Wu JC, Hataishi R, Aretz HT, Picard MH et al (2004) Pressure overload-induced LV hypertrophy and dysfunction in mice are exacerbated by congenital NOS3 deficiency. Am J Physiol 286:H1070–H1075
- Ignarro LJ (1990) Biosynthesis and metabolism of endotheliumderived nitric oxide. Annu Rev Pharmacol Toxicol 30:535–560
- 22. Janssens S, Pokreisz P, Schoonjans L, Pellens M, Vermeersch P, Tjwa M et al (2004) Cardiomyocyte-specific overexpression of nitric oxide synthase 3 improves left ventricular performance and reduces compensatory hypertrophy after myocardial infarction. Circ Res 94:1256–1262
- Jones SP, Greer JJ, van Haperen R, Duncker DJ, de Crom R, Lefer DJ (2003) Endothelial nitric oxide synthase overexpression attenuates congestive heart failure in mice. Proc Natl Acad Sci USA 100:4891–4896
- 24. Kadowaki T, Yamauchi T, Kubota N, Hara K, Ueki K, Tobe K (2006) Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. J Clin Invest 116:1784–1792
- 25. Kawashima S, Yamashita T, Ozaki M, Ohashi Y, Azumi H, Inoue N et al (2001) Endothelial NO synthase overexpression inhibits lesion formation in mouse model of vascular remodeling. Arterioscler Thromb Vasc Biol 21:201–207
- 26. Knowles JW, Reddick RL, Jennette JC, Shesely EG, Smithies O, Maeda N (2000) Enhanced atherosclerosis and kidney dysfunction in eNOS<sup>-/-</sup>Apoe<sup>-/-</sup> mice are ameliorated by enalapril treatment. J Clin Invest 105:451–458
- Koglin J, Glysing-Jensen T, Mudgett JS, Russell ME (1998) Exacerbated transplant arteriosclerosis in inducible nitric oxidedeficient mice. Circulation 97:2059–2065
- 28. Kohro T, Hayashi D, Okada Y, Yamazaki T, Nagai R (2008) Demographics and changes in medical/interventional treatment of

coronary artery disease patients over a 3.5-year period in Japan: The Japanese Coronary Artery Disease Study: Trend examination. Circ J 72:1397–1402

- 29. Kuhlencordt PJ, Chen J, Han F, Astern J, Huang PL (2001) Genetic deficiency of inducible nitric oxide synthase reduces atherosclerosis and lowers plasma lipid peroxides in apolipoprotein E-knockout mice. Circulation 103:3099–3104
- 30. Kuhlencordt PJ, Gyurko R, Han F, Scherrer-Crosbie M, Aretz TH, Hajjar R et al (2001) Accelerated atherosclerosis, aortic aneurysm formation, and ischemic heart disease in apolipoprotein E/endothelial nitric oxide synthase double-knockout mice. Circulation 104:448–454
- Kuhlencordt PJ, Hotten S, Schodel J, Rutzel S, Hu K, Widder J et al (2006) Atheroprotective effects of neuronal nitric oxide synthase in apolipoproteine knockout mice. Arterioscler Thromb Vasc Biol 26:1539–1544
- 32. Laine P, Kaartinen M, Penttila A, Panula P, Paavonen T, Kovanen PT (1999) Association between myocardial infarction and the mast cells in the adventitia of the infarct-related coronary artery. Circulation 99:361–369
- Lamping KG, Nuno DW, Shesely EG, Maeda N, Faraci FM (2000) Vasodilator mechanisms in the coronary circulation of endothelial nitric oxide synthase-deficient mice. Am J Physiol 279:H1906–H1912
- 34. Landmesser U, Engberding N, Bahlmann FH, Schaefer A, Wiencke A, Heineke A et al (2004) Statin-induced improvement of endothelial progenitor cell mobilization, myocardial neovascularization, left ventricular function, and survival after experimental myocardial infarction requires endothelial nitric oxide synthase. Circulation 110:1933–1939
- Laubach VE, Shesely EG, Smithies O, Sherman PA (1995) Mice lacking inducible nitric oxide synthase are not resistant to lipopolysaccharideinduced death. Proc Natl Acad Sci USA 92:10688–10692
- 36. Liu YH, Xu J, Yang XP, Yang F, Shesely E, Carretero OA (2002) Effect of ACE inhibitors and angiotensin II type 1 receptor antagonists on endothelial NO synthase knockout mice with heart failure. Hypertension 39:375–381
- 37. Loyer X, Gomez AM, Milliez P, Fernandez-Velasco M, Vangheluwe P, Vinet L et al (2008) Cardiomyocyte overexpression of neuronal nitric oxide synthase delays transition toward heart failure in response to pressure overload by preserving calcium cycling. Circulation 117:3187–3198
- MacMicking JD, Nathan C, Hom G, Chartrain N, Fletcher DS, Trumbauer M et al (1995) Altered responses to bacterial infection and endotoxic shock in mice lacking inducible nitric oxide synthase. Cell 81:641–650
- Massion PB, Balligand JL (2003) Modulation of cardiac contraction, relaxation and rate by the endothelial nitric oxide synthase (eNOS): Lessons from genetically modified mice. J Physiol 546:63–75
- 40. Matoba T, Shimokawa H, Nakashima M, Hirakawa Y, Mukai Y, Hirano K, Kanaide H, Takeshita A (2000) Hydrogen peroxide is an endothelium-derived hyperpolarizing factor in mice. J Clin Invest 106:1521–1530
- Matsuzawa Y, Funahashi T, Kihara S, Shimomura I (2004) Adiponectin and metabolic syndrome. Arterioscler Thromb Vasc Biol 24:29–33
- 42. Morikawa K, Shimokawa H, Matoba T, Kubota H, Akaike T, Talukder MA, Hatanaka M, Fujiki T, Maeda H, Takahashi S, Takeshita A (2003) Pivotal role of Cu, Zn-superoxide dismutase in endothelium-dependent hyperpolarization. J Clin Invest 112:1871–1879
- Morishita T, Tsutsui M, Shimokawa H, Horiuchi M, Tanimoto A, Suda O et al (2002) Vasculoprotective roles of neuronal nitric oxide synthase. FASEB J 16:1994–1996

- 44. Morishita T, Tsutsui M, Shimokawa H, Sabanai K, Tasaki H, Suda O et al (2005) Nephrogenic diabetes insipidus in mice lacking all nitric oxide synthase isoforms. Proc Natl Acad Sci USA 102:10616–10621
- 45. Moroi M, Zhang L, Yasuda T, Virmani R, Gold HK, Fishman MC et al (1998) Interaction of genetic deficiency of endothelial nitric oxide, gender, and pregnancy in vascular response to injury in mice. J Clin Invest 101:1225–1232
- 46. Mungrue IN, Gros R, You X, Pirani A, Azad A, Csont T et al (2002) Cardiomyocyte overexpression of iNOS in mice results in peroxynitrite generation, heart block, and sudden death. J Clin Invest 109:735–743
- Mungrue IN, Husain M, Stewart DJ (2002) The role of NOS in heart failure: lessons from murine genetic models. Heart Fail Rev 7:407–422
- 48. Murad F (1997) What are the molecular mechanisms for the antiproliferative effects of nitric oxide and cGMP in vascular smooth muscle? Circulation 95:1101–1103
- 49. Nakamura T, Tsubono Y, Kameda-Takemura K, Funahashi T, Yamashita S, Hisamichi S et al (2001) Magnitude of sustained multiple risk factors for ischemic heart disease in Japanese employees: a case-control study. Jpn Circ J 65:11–17
- Nakata S, Tsutsui M, Shimokawa H, Suda O, Morishita T, Shibata K et al (2008) Spontaneous myocardial infarction in mice lacking all nitric oxide synthase isoforms. Circulation 117:2211–2223
- 51. Nakata S, Tsutsui M, Shimokawa H, Tamura M, Tasaki H, Morishita T et al (2005) Vascular neuronal NO synthase is selectively upregulated by platelet-derived growth factor. Arterioscler Thromb Vasc Biol 25:2502–2508
- 52. Nakata S, Tsutsui M, Shimokawa H, Yamashita T, Tanimoto A, Tasaki H et al (2007) Statin treatment upregulates vascular neuronal nitric oxide synthase through Akt/NF-κB pathway. Arterioscler Thromb Vasc Biol 27:92–98
- 53. Ohashi Y, Kawashima S, Hirata K, Yamashita T, Ishida T, Inoue N et al (1998) Hypotension and reduced nitric oxide-elicited vasorelaxation in transgenic mice overexpressing endothelial nitric oxide synthase. J Clin Invest 102:2061–2071
- 54. Ozaki M, Kawashima S, Yamashita T, Hirase T, Namiki M, Inoue N et al (2002) Overexpression of endothelial nitric oxide synthase accelerates atherosclerotic lesion formation in apoE-deficient mice. J Clin Invest 110:331–340
- Packer MA, Hemish J, Mignone JL, John S, Pugach I, Enikolopov G (2005) Transgenic mice overexpressing nNOS in the adult nervous system. Cell Mol Biol 51:269–277
- 56. Packer MA, Stasiv Y, Benraiss A, Chmielnicki E, Grinberg A, Westphal H, Goldman SA, Enikolopov G (2003) Nitric oxide negatively regulates mammalian adult neurogenesis. Proc Natl Acad Sci USA 100:9566–9571
- Park CS, Park R, Krishna G (1996) Constitutive expression and structural diversity of inducible isoform of nitric oxide synthase in human tissues. Life Sci 59:219–225
- Prabhu SD (2004) Nitric oxide protects against pathological ventricular remodeling: Reconsideration of the role of NO in the failing heart. Circ Res 94:1155–1157
- Rudic RD, Shesely EG, Maeda N, Smithies O, Segal SS, Sessa WC (1998) Direct evidence for the importance of endotheliumderived nitric oxide in vascular remodeling. J Clin Invest 101:731–736
- 60. Sam F, Sawyer DB, Xie Z, Chang DL, Ngoy S, Brenner DA et al (2001) Mice lacking inducible nitric oxide synthase have improved left ventricular contractile function and reduced apoptotic cell death late after myocardial infarction. Circ Res 89:351– 356
- 61. Saraiva RM, Hare JM (2006) Nitric oxide signaling in the cardiovascular system: implications for heart failure. Curr Opin Cardiol 21:221–228

- 62. Saraiva RM, Minhas KM, Raju SV, Barouch LA, Pitz E, Schuleri KH et al (2005) Deficiency of neuronal nitric oxide synthase increases mortality and cardiac remodeling after myocardial infarction: role of nitroso-redox equilibrium. Circulation 112: 3415–3422
- Scherrer-Crosbie M, Ullrich R, Bloch KD, Nakajima H, Nasseri B, Aretz HT et al (2001) Endothelial nitric oxide synthase limits left ventricular remodeling after myocardial infarction in mice. Circulation 104:1286–1291
- 64. Shesely EG, Maeda N, Kim HS, Desai KM, Krege JH, Laubach VE et al (1996) Elevated blood pressures in mice lacking endothelial nitric oxide synthase. Proc Natl Acad Sci USA 93:13176–13181
- Shimokawa H (1999) Primary endothelial dysfunction: atherosclerosis. J Mol Cell Cardiol 31:23–37
- 66. Shimokawa H, Morikawa K (2005) Hydrogen peroxide is an endothelium-derived hyperpolarizing factor in animals and humans. J Mol Cell Cardiol 39:725–732
- 67. Shioji K, Moriwaki S, Takeuchi Y, Uegaito T, Mutsuo S, Matsuda M (2007) Relationship of serum adiponectin level to adverse cardiovascular events in patients who undergo percutaneous coronary intervention. Circ J 71:675–680
- 68. Son H, Hawkins RD, Martin K, Kiebler M, Huang PL, Fishman MC et al (1996) Long-term potentiation is reduced in mice that are doubly mutant in endothelial and neuronal nitric oxide synthase. Cell 87:1015–1023
- 69. Takaki A, Morikawa K, Tsutsui M, Murayama Y, Takes E, Yamagishi H, Ohashi J, Yada T, Yanagihara N, Shimokawa H (2008) Crucial role of nitric oxide synthases system in endotheliumdependent hyperpolarization in mice. J Exp Med 205:2053– 2063
- 70. Takamura T, Kato I, Kimura N, Nakazawa T, Yonekura H, Takasawa S et al (1998) Transgenic mice overexpressing type 2 nitric-oxide synthase in pancreatic beta cells develop insulindependent diabetes without insulitis. J Biol Chem 273:2493– 2496
- 71. Takeno M, Yasuda S, Otsuka Y, Morii I, Kawamura A, Yano K et al (2008) Impact of metabolic syndrome on the long-term survival of patients with acute myocardial infarction: potential association with C-reactive protein. Circ J 72:415–419
- Tranguch S, Huet-Hudson Y (2003) Decreased viability of nitric oxide synthase double knockout mice. Mol Reprod Dev 65:175– 179
- Tsutsui M (2004) Neuronal nitric oxide synthase as a novel antiatherogenic factor. J Atheroscler Thromb 11:41–48
- Tsutsui M, Nakata S, Shimokawa H, Otsuji Y, Yanagihara N (2008) Spontaneous myocardial infarction and nitric oxide synthase. Trends Cardiovasc Med 18:275–279
- 75. Tsutsui M, Shimokawa H, Morishita T, Nakashima Y, Yanagihara N (2006) Development of genetically engineered mice lacking all three nitric oxide synthases. J Pharmacol Sci 102:147– 154
- Tsutsui M, Shimokawa H, Otsuji Y, Ueta Y, Sasaguri Y, Yanagihara N (2009) Nitric oxide synthases and cardiovascular diseases. Insights from genetically modified mice. Circ J 73:986– 993
- 77. van Haperen R, de Waard M, van Deel E, Mees B, Kutryk M, van Aken T et al (2002) Reduction of blood pressure, plasma cholesterol, and atherosclerosis by elevated endothelial nitric oxide. J Biol Chem 277:48803–48807
- Vanhoutte PM (2009) Endothelial dysfunction. The first step toward coronary arteriosclerosis. Circ J 73:595–601
- Vanhoutte PM, Shimokawa H (1989) Endothelium-derived relaxing factor and coronary vasospasm. Circulation 80:1–9
- Vasquez-Vivar J, Kalyanaraman B, Martasek P, Hogg N, Masters BS, Karoui H et al (1998) Superoxide generation by endothelial

nitric oxide synthase: the influence of cofactors. Proc Natl Acad Sci USA 95:9220–9225

- Wang W, Wang S, Yan L, Madara P, Del Pilar CA, Wesley RA et al (2000) Superoxide production and reactive oxygen species signaling by endothelial nitric-oxide synthase. J Biol Chem 275:16899–16903
- 82. Ward ME, Toporsian M, Scott JA, Teoh H, Govindaraju V, Quan A et al (2005) Hypoxia induces a functionally significant and translationally efficient neuronal NO synthase mRNA variant. J Clin Invest 115:3128–3139
- Wei XQ, Charles IG, Smith A, Ure J, Feng GJ, Huang FP et al (1995) Altered immune responses in mice lacking inducible nitric oxide synthase. Nature 375:408–411
- 84. Wilcox JN, Subramanian RR, Sundell CL, Tracey WR, Pollock JS, Harrison DG et al (1997) Expression of multiple isoforms of nitric oxide synthase in normal and atherosclerotic vessels. Arterioscler Thromb Vasc Biol 17:2479–2488
- 85. Yogo K, Shimokawa H, Funakoshi H, Kandabashi T, Miyata K, Okamoto S et al (2000) Different vasculoprotective roles of NO synthase isoforms in vascular lesion formation in mice. Arterioscler Thromb Vasc Biol 20:E96–E100
- 86. Zhang P, Xu X, Hu X, van Deel ED, Zhu G, Chen Y (2007) Inducible nitric oxide synthase deficiency protects the heart from systolic overload-induced ventricular hypertrophy and congestive heart failure. Circ Res 100:1089–1098