

## Chapter 2

# Pathophysiology and Molecular Mechanisms of Coronary Artery Spasm



Kimio Satoh and Hiroaki Shimokawa

**Abstract** Rho-kinase plays a central role in the pathogenesis of coronary artery spasm caused by vascular smooth muscle cell (VSMC) hypercontraction. Rho-kinase belongs to the family of serine/threonine kinases and is an important downstream effector of the small GTP-binding protein RhoA. Two isoforms of Rho-kinase, ROCK1 and ROCK2, have different functions with ROCK1 for circulating inflammatory cells and ROCK2 for vascular smooth muscle cells. The RhoA/Rho-kinase pathway plays an important role in many cellular functions, including contraction, motility, proliferation, and apoptosis, leading to the development of cardiovascular diseases. In addition to vasospasm, important roles of Rho-kinase *in vivo* have been demonstrated in the pathogenesis of arteriosclerosis, ischemia/reperfusion injury, hypertension, pulmonary hypertension, stroke, and heart failure. Furthermore, the beneficial effects of fasudil, a selective Rho-kinase inhibitor, have been demonstrated for the treatment of several cardiovascular diseases in animals and humans. Thus, the Rho-kinase pathway is an important new therapeutic target in vasospasm and other cardiovascular diseases.

**Keywords** Vasospasm · Rho-kinase · Cardiovascular disease · Oxidative stress · Small G proteins

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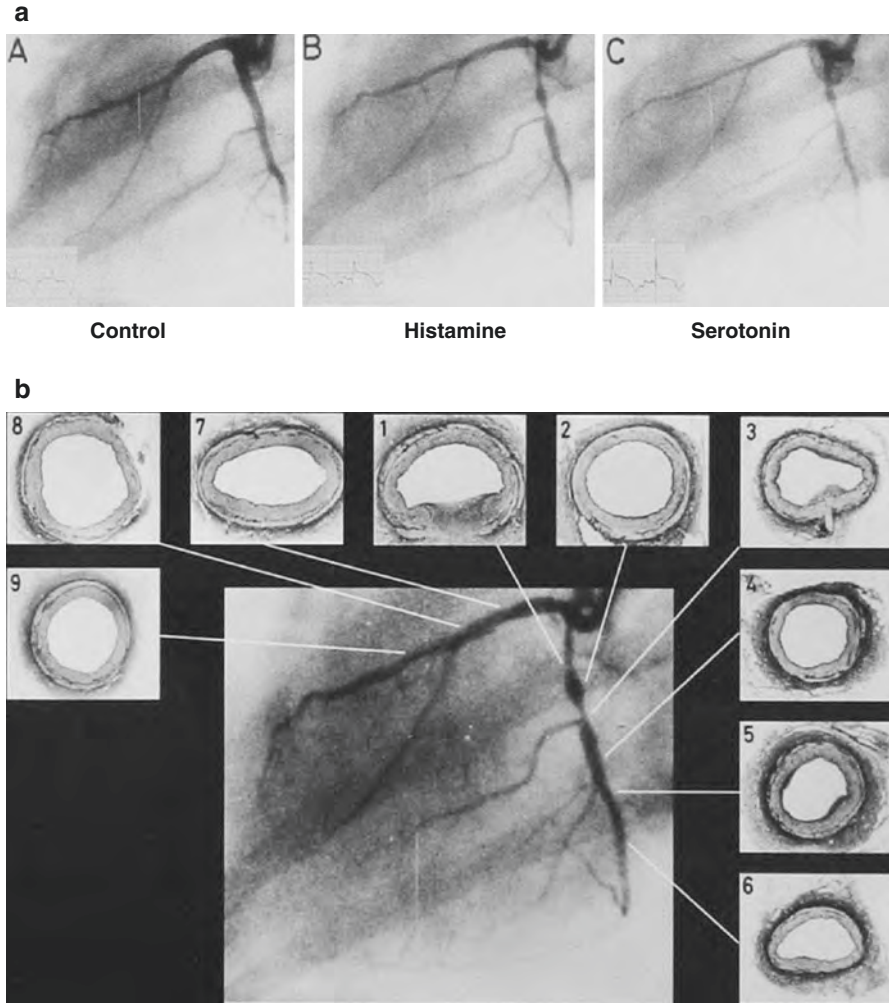
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## 2.1 Development of Animal Models of Coronary Artery Spasm and Identification of Important Pathogenetic Roles of Rho-Kinase

Rho-kinase activation plays a central role in the pathogenesis of coronary artery spasm caused by vascular smooth muscle cell (VSMC) hypercontraction. In an animal model in pigs *in vivo*, we examined whether atherosclerotic coronary lesion, induced by a combination of balloon endothelium removal and high-cholesterol feeding, exhibits hyperresponsiveness to vasoconstrictor agents [1]. Importantly, intracoronary administration of serotonin induced coronary artery spasm at the atherosclerotic lesion, and there was a close topological correlation between the spastic site and atherosclerotic lesion (Fig. 2.1a, b) [1]. This is the first experimental evidence for the close relationship between coronary artery spasm and coronary atherosclerosis [1]. Next, we further examined whether chronic adventitial inflammation could cause vasospastic activity of the coronary artery without endothelium removal in pigs. Two weeks after the adventitial application of interleukin-1 $\beta$  (IL-1 $\beta$ ), coronary angiography showed the development of mild stenotic lesion, where intracoronary administration of serotonin repeatedly caused coronary spasm (Fig. 2.1c) [2]. Histological examination showed adventitial accumulation of inflammatory cells, mild neointimal formation, and a marked reduction in vascular cross-sectional area (Fig. 2.1d) [2]. These results provided the first experimental evidence for the role of adventitial inflammation in the pathogenesis of coronary artery spasm. Delayed cerebral ischemia due to cerebral vasospasm remains a major cause of morbidity in patients with subarachnoid hemorrhage (SAH). It has been demonstrated that Rho-kinase is substantially involved in the pathogenesis of cerebral vasospasm after SAH [3]. Coronary artery spasm plays an important role in variant angina, myocardial infarction, and sudden cardiac death [4]. It was demonstrated that elevated serum level of cortisol, one of the important stress hormones, causes coronary hyperreactivity through activation of Rho-kinase in pigs *in vivo* [5]. The activity and the expression of ROCKs are enhanced at the inflammatory/arteriosclerotic coronary lesions [6]. Accumulating evidence indicates that Rho-kinase plays a crucial role in the pathogenesis of coronary artery spasm. Intracoronary administration of fasudil [7] and of hydroxyfasudil [8] inhibited coronary spasm in pigs *in vivo* [2]. We have demonstrated that fasudil is effective in preventing coronary spasm and resultant myocardial ischemia in patients with vasospastic angina [9]. Thus, fasudil is useful for the treatment of ischemic coronary syndromes caused by coronary artery spasm. Fasudil is also effective in treating patients with microvascular angina [10]. The clinical trials for the effects of fasudil in Japanese patients with stable-effort angina demonstrated that the long-term oral treatment with the Rho-kinase inhibitor is effective in ameliorating exercise tolerance in those patients [11]. We also have recently demonstrated that Rho-kinase activity in circulating neutrophils is an useful biomarker for the diagnosis and disease activity assessment in patients with VSA [12].



**Fig. 2.1** Coronary artery spasm induced in two porcine models in vivo. **(a, b)** Coronary artery spasm was induced in atherosclerotic miniature pigs induced by balloon endothelial injury and high-cholesterol feeding **(a)**, where topological correlation was noted between the spastic sites and the early atherosclerotic lesions **(b)**. **(c, d)** Coronary artery spasm was induced in pigs with adventitial inflammation **(c)**, where intimal thickening and negative remodeling were noted **(d)**. (Reproduced from Shimokawa et al. [1, 2])

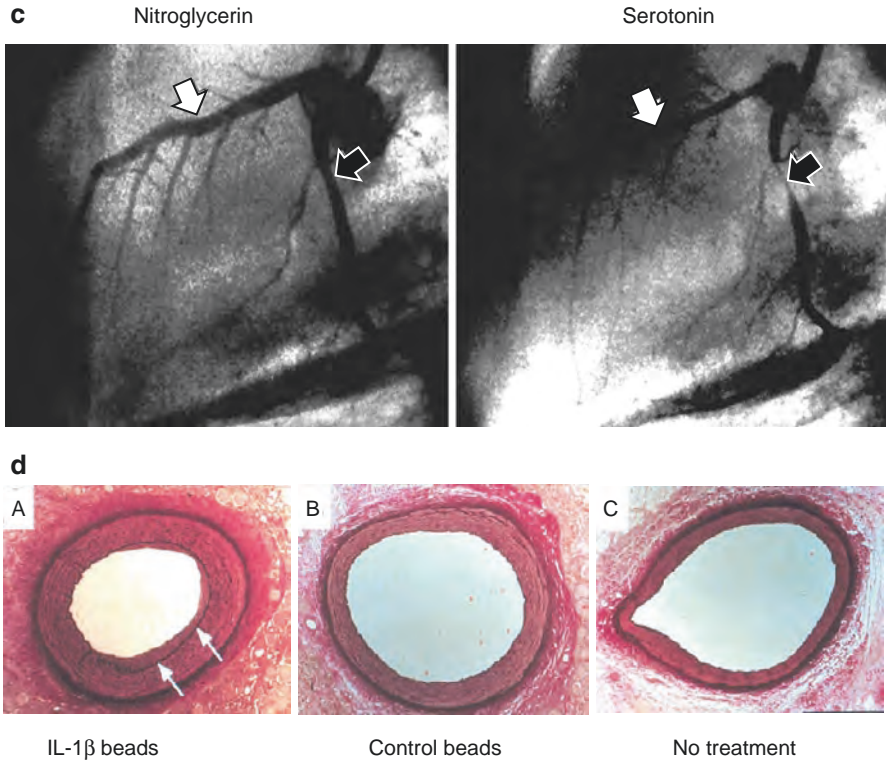
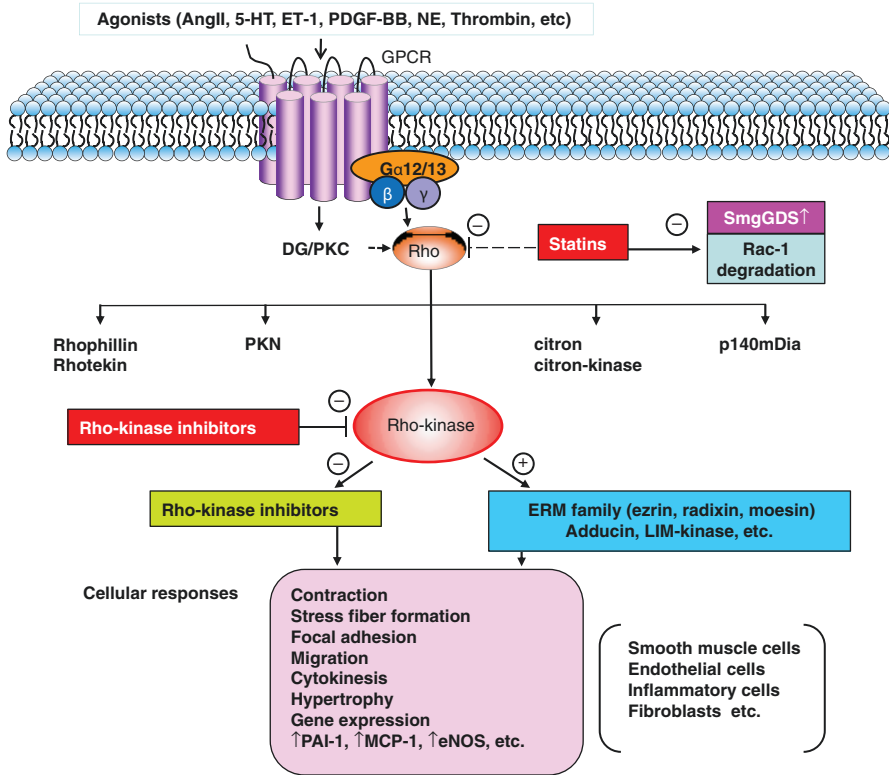


Fig. 2.1 (continued)

## 2.2 The Rho/Rho-Kinase System in Vascular Contraction

Rho-kinase belongs to the family of serine/threonine kinases and is an important downstream effector of the small GTP-binding protein RhoA. The Rho family of small G proteins comprises 20 members of ubiquitously expressed proteins in mammals, including RhoA, Rac1, and Cdc42 [13–15]. Among them, RhoA is the best-characterized protein that acts as a molecular switch that cycles between an inactive GDP-bound and an active GTP-bound conformation interacting with downstream targets to elicit a variety of cellular responses (Fig. 2.2) [16]. The activity of RhoA is controlled by the guanine nucleotide exchange factors (GEFs) that catalyze exchange of GDP for GTP [17]. In contrast, GTPase activating proteins (GAPs) stimulate the intrinsic GTPase activity and inactivate RhoA [18]. Additionally, it has been demonstrated that guanine nucleotide dissociation inhibitors (GDIs) block spontaneous RhoA activation (Fig. 2.2) [19].

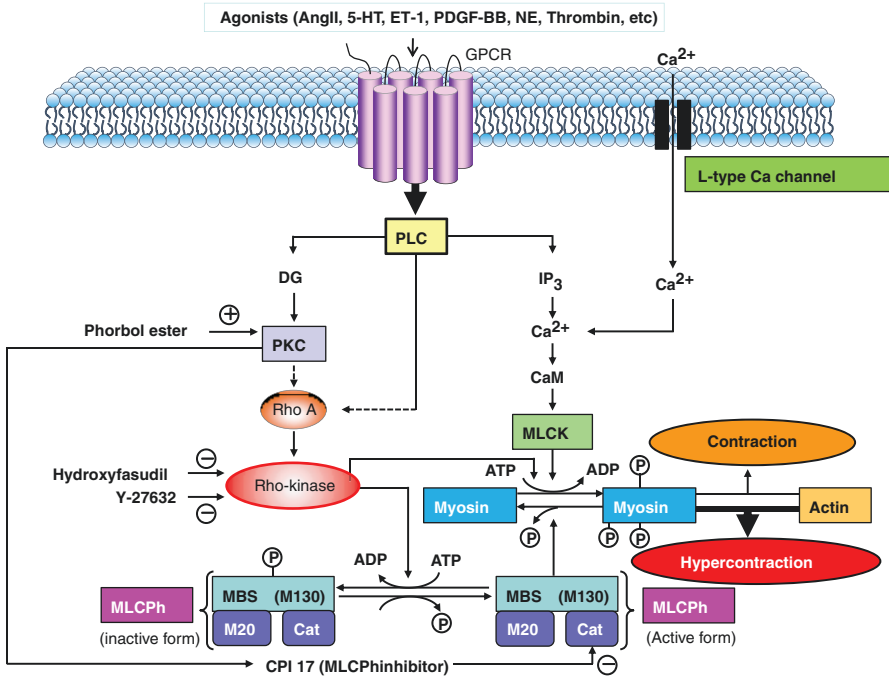
In 1996, Rho-kinase (Rho-kinase  $\alpha$ /ROCK 2/ROK $\alpha$  and Rho-kinase  $\beta$ /ROCK 1/ROK $\beta$ ) was identified as the effector of Rho (Fig. 2.2) [20–22]. Phosphorylation of



**Fig. 2.2** The important roles of Rho/Rho-kinase pathway in the pathogenesis of cardiovascular diseases. The Rho/Rho-kinase pathway plays important roles in the pathogenesis of vasospastic disorders as well as atherosclerotic cardiovascular diseases in general. (Reproduced from Shimokawa et al. [27])

myosin light chain (MLC) is a key event in the regulation of VSMC contraction (Fig. 2.3). MLC is phosphorylated by  $Ca^{2+}$ -calmodulin-activated MLC kinase (MLCK) and dephosphorylated by MLC phosphatase (MLCP) (Fig. 2.3). Agonists bind to G-protein-coupled receptors and induce contraction by increasing both cytosolic  $Ca^{2+}$  concentration and Rho-kinase activity through mediating GEF. The substrates of Rho-kinase have been identified, including MLC, myosin-binding subunit (MBS) or myosin phosphatase target subunit (MYPT-1), ERM family, adducin, PTEN, and LIM-kinases (Figs. 2.2 and 2.3). Rho-kinase enhances MLC phosphorylation through inhibition of MBS of myosin phosphatase and mediate agonists-induced VSMC contraction (Fig. 2.3).

The interaction between endothelial cells (ECs) and VSMCs plays an important role in regulating vascular integrity and vascular homeostasis [23, 24]. ECs release vasoactive factors, such as prostacyclin, nitric oxide (NO) and endothelium-derived hyperpolarizing factor (EDHF), participating in the regulation of vascular tone and arterial resistance [1, 25–27]. It has been demonstrated that both endothelial NO



**Fig. 2.3** Molecular mechanisms of vascular smooth muscle cells hypercontraction for coronary spasm. The central molecular mechanism of vascular smooth muscle cell hypercontraction for coronary spasm is Rho-kinase-mediated enhancement of myosin light chain phosphorylations through inhibition of myosin light chain phosphatase. (Reproduced from Shimokawa et al. [27])

production and NO-mediated signaling in VSMCs are targets and effectors of the RhoA/Rho-kinase pathway [23, 28]. In ECs, the RhoA/Rho-kinase pathway negatively regulates NO production [29]. In contrast, VSMCs are among the most plastic of all cells in their ability to respond to different stimuli [30–32]. The initial works in our laboratory on the therapeutic importance of Rho-kinase were previously summarized [23, 33]. Since then, a significant progress has been made in our knowledge on the therapeutic importance of Rho-kinase in cardiovascular medicine. In this article, we will briefly review the recent progress in the translational research on the therapeutic importance of the Rho-kinase pathway in cardiovascular medicine.

### 2.3 Substrates of Rho-Kinase

Rho-kinase is a serine/threonine kinase with a molecular weight of ~160 kDa. Two isoforms of Rho-kinase encoded by 2 different genes have been identified [34–36]. In humans, ROCK1 and ROCK2 genes are located separately on chromosome 18 and chromosome 2, respectively. They are ubiquitously expressed in invertebrates



and vertebrates with ROCK1 especially in circulating inflammatory cells and ROCK2 in VSMCs. ROCKs consist of 3 major domains, including a kinase domain in its N-terminal domain, a coiled-coil domain that includes Rho-binding domain in its middle portion, and a putative pleckstrin homology (PH) domain in its C-terminal domain [13]. Rho-kinase activity is enhanced by binding of GTP-bound active form of RhoA [35] (Fig. 2.2). Rho-kinase inhibitors, fasudil [8] and Y-27632 [37], have been developed and they inhibit Rho-kinase activity in a competitive manner with ATP at the Rho-binding site [38]. It has been demonstrated that hydroxyfasudil, a major active metabolite of fasudil, exerts a more specific inhibitory effect on Rho-kinase [8, 39].

Although regulation of Rho-kinase expression has not been fully elucidated, some studies have reported changes in Rho-kinase expression. Functional differences between ROCK1 and ROCK2 have been reported; ROCK1 is specifically cleaved by caspase-3, whereas ROCK2 is cleaved by granzyme B [40, 41]. The small G-protein RhoE specially binds to the N-terminal region of ROCK1 at the kinase domain, whereas the MYPT1 binds specially ROCK2 [42, 43]. RhoE binding to ROCK1 inhibits its activity and prevents RhoA binding to the Rho-binding domain [44]. Both ROCK1 and ROCK2 mRNAs and proteins are upregulated by angiotensin II (AngII) via AT1 receptor stimulation and by interleukin-1 $\beta$  (IL-1 $\beta$ ) [45]. A number of Rho-kinase substrates have been identified [46] (Fig. 2.2) and Rho-kinase-mediated substrate phosphorylation causes actin filament formation, organization, and cytoskeleton rearrangement (Fig. 2.2) [47]. The N-terminal regions, upstream of the kinase domains of Rho-kinase, may play a role in determining substrate specificity of the 2 isoforms [47].

The majority of Rho-kinase substrates have been identified *in vitro*. Thus, ROCK1- and ROCK2-deficient mice have been generated to further elucidate the functions of the ROCK isoforms [48, 49]. Importantly, ROCK1-deficient mice showed the eyelids opened at birth [49], whereas ROCK2-deficient mice placental dysfunction and fetal death [48, 50–52]. Thus, the role of ROCK2, the main isoform in the cardiovascular system, remained to be fully elucidated *in vivo*. In order to address this point, we have recently developed VSMC-specific ROCK2-deficient mice and found the crucial role of ROCK2 in the development of hypoxia-induced pulmonary hypertension [30].

## 2.4 Rho-Kinase-Mediated Inflammation and Oxidative Stress

Rho-kinase augments inflammation by inducing pro-inflammatory molecules, including IL-6 [53], monocyte chemoattractant protein (MCP)-1 [54], macrophage migration inhibitory factor (MIF) [55, 56], and sphingosine-1-phosphate (S1P) [57]. In ECs, Rho-kinase downregulates eNOS [58] and substantially activates pro-inflammatory pathways including enhanced expression of adhesion molecules. The expression of Rho-kinase is accelerated by inflammatory stimuli, such as AngII and

IL-1 $\beta$  [45], and by remnant lipoproteins in human coronary VSMCs [59]. Rho-kinase also upregulates NAD(P)H oxidases and augments AngII-induced ROS production [39]. Several growth factors are known to be secreted from VSMC in response to oxidative stress. Rho GTPases including RhoA are key regulators in signaling pathways linked to actin cytoskeletal rearrangement [60]. RhoA plays a central role in vesicular trafficking pathways by controlling organization of actin cytoskeleton. It has been reported that active participation of Rho GTPases is required for secretion. Myosin II is involved in secretory mechanisms as a motor for vesicle transport [61]. Rho-kinase mediates myosin II activation via phosphorylation and inactivation of myosin II light chain phosphatase [20]. Thus, the Rho/Rho-kinase is important for the secretion of inflammatory cytokines and growth factors (Fig. 2.2).

## 2.5 Rho-Kinase in Vascular Function and Contraction

Rho-kinase has been implicated in the pathogenesis of cardiovascular disease, in part by promoting VSMC proliferation [62–64]. Changes in vascular redox state are a common pathway involved in the pathogenesis of vasospastic angina (VSA), atherosclerosis, aortic aneurysms, and vascular stenosis. Vascular ROS formation can be stimulated by mechanical stretch, pressure overload, shear stress, environmental factors (e.g. hypoxia), and growth factors (e.g. AngII) [65]. Importantly, Rho-kinase is substantially involved in the vascular effects of various vasoactive factors, including AngII [39, 54, 66, 67], thrombin [68, 69], platelet-derived growth factor [70], extracellular nucleotides [71], and urotensin [72] (Fig. 2.2). It has previously been shown that statins enhance eNOS mRNA by cholesterol-independent mechanisms involving inhibition of Rho geranylgeranylation [73]. Rho-kinase plays an important role in mediating various cellular functions, not only VSMC contraction [74, 75] but also actin cytoskeleton organization [76, 77], adhesion, and cytokinesis [33]. Thus, Rho-kinase plays a crucial role in the development of cardiovascular disease through ROS production, inflammation, EC damage, VSMC contraction and proliferation (Figs. 2.2 and 2.3).

## 2.6 Rho-Kinase in Arteriosclerosis

As mentioned above, Rho-kinase plays a crucial role in the ROS augmentation and vascular inflammation. ROS have been implicated in the pathogenesis of neointima formation in part by promoting VSMC growth [64, 78] and by stimulating pro-inflammatory events [79–81]. Accumulating evidence indicates that Rho-kinase inhibitors have broad pharmacological properties [33, 75, 82]. The beneficial effects of long-term inhibition of Rho-kinase for the treatment of cardiovascular disease



have been demonstrated in various animal models, such as coronary artery spasm, arteriosclerosis, restenosis, ischemia/reperfusion injury, hypertension, pulmonary hypertension, stroke, and cardiac hypertrophy/heart failure [33, 75, 82]. Gene transfer of dominant-negative Rho-kinase reduced the neointimal formation in pigs [83]. Long-term treatment with a Rho-kinase inhibitor suppressed neointima formation after vascular injury in vivo [84, 85], MCP-1-induced vascular lesion formation [86], constrictive remodeling [87], in-stent restenosis [88], and the development of cardiac allograft vasculopathy [56] (Fig. 2.2).

Arteriosclerosis is a slowly progressing inflammatory process of the arterial wall that involves the intima, media, and adventitia [33, 75]. Accumulating evidence indicates that Rho-kinase-mediated pathway is substantially involved in EC dysfunction [58, 69], VSMC contraction [89], VSMC proliferation and migration in the media [90], and accumulation of inflammatory cells in the adventitia [86]. Those Rho-kinase-mediated cellular responses lead to the development of vascular disease. In fact, mRNA expression of ROCKs is enhanced at the inflammatory and arteriosclerotic arterial lesions in animals [89] and humans [91]. In the context of atherosclerosis, Rho-kinase should be regarded as a pro-inflammatory and pro-atherogenic molecule. Thus, Rho-kinase is an important new therapeutic target for the treatment of atherosclerosis (Fig. 2.2).

## 2.7 Rho-Kinase in Myocardial Ischemia and Heart Failure

ROS production and Rho-kinase activation play a crucial role in myocardial damage after ischemia/reperfusion. Consistently, we have demonstrated that pretreatment with fasudil before reperfusion prevents endothelial dysfunction and reduces myocardial infarction size in dogs in vivo [92]. The beneficial effect of fasudil has also been demonstrated in a rabbit model of myocardial ischemia induced by intravenous administration of endothelin-1 [93], a canine model of pacing-induced myocardial ischemia [94], and a rat model of vasopressin-induced chronic myocardial ischemia [95]. AngII plays a key role in many physiological and pathological processes in cardiac cells, including cardiac hypertrophy [96]. Understanding the molecular mechanisms for AngII-induced myocardial disorders is important to develop new therapies for cardiac dysfunction [97]. One important mechanism now recognized to be involved in AngII-induced cardiac hypertrophy is ROS production [98, 99], however, the precise mechanism by which ROS cause myocardial hypertrophy and dysfunction still remains to be fully elucidated [100]. It has been demonstrated that cardiac troponin is a substrate of Rho-kinase [101]. Rho-kinase phosphorylates troponin and inhibits tension generation in cardiomyocytes. We have demonstrated that Rho-kinase inhibition with fasudil suppresses the development of cardiac hypertrophy and diastolic heart failure in Dahl salt-sensitive rats [102]. In patients with heart failure, intra-arterial infusion of fasudil caused preferential increase in forearm blood flow as compared with control subjects, suggesting an involvement of

Rho-kinase in the increased peripheral vascular resistance in patients with heart failure [103].

## 2.8 Rho-Kinase in Hypertension and Pulmonary Hypertension

Short-term administration of Y-27632, another Rho-kinase inhibitor, preferentially reduces systemic blood pressure in a dose-dependent manner in rat models of systemic hypertension, suggesting an involvement of Rho-kinase in the pathogenesis of hypertension [37]. The expression of Rho-kinase was significantly increased in spontaneously hypertensive rats (SHR) [104]. Local administration of a small amount of hydroxyfasudil into the nucleus tractus solitarii of the brain stem causes sustained decrease in heart rate and blood pressure in SHR but not in normotensive rats, suggesting that Rho-kinase is involved in the central mechanisms of sympathetic nerve activity [105]. Inhibition of Rho-kinase in the brain stem also augments baroreflex control of heart rate in rats [106]. Pulmonary hypertension (PH) is associated with hypoxic exposure, endothelial dysfunction, VSMC hypercontraction and proliferation, enhanced ROS production, and inflammatory cell migration, for which Rho-kinase may also be substantially involved. Indeed, long-term treatment with fasudil suppresses the development of monocrotaline-induced PH in rats [107] and of hypoxia-induced PH in mice [108]. Recently, we were able to obtain direct evidence for Rho-kinase activation in patients with pulmonary arterial hypertension (PAH) [109]. Furthermore, intravenous infusion of fasudil significantly reduced pulmonary vascular resistance in patients with PAH, indicating an involvement of Rho-kinase in the pathogenesis of PAH in humans [110].

## 2.9 Conclusions

Accumulating evidence has indicated that Rho-kinase plays important roles in the pathogenesis of a wide range of cardiovascular diseases in general and coronary vasomotion abnormalities in particular. Additionally, Rho-kinase inhibitors are useful for the treatment of those cardiovascular diseases. In conclusion, accumulating experimental and clinical evidence indicates that Rho-kinase is an important new target for the treatment of VSA and cardiovascular diseases.

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## References

1. Shimokawa H, Tomoike H, Nabeyama S, Yamamoto H, Araki H, Nakamura M, Ishii Y, Tanaka K. Coronary artery spasm induced in atherosclerotic miniature swine. *Science*. 1983;221(4610):560–2. <https://doi.org/10.1126/science.6408736>.
2. Shimokawa H, Ito A, Fukumoto Y, Kadokami T, Nakaïke R, Sakata M, Takayanagi T, Egashira K, Takeshita A. Chronic treatment with interleukin-1 $\beta$  induces coronary intimal lesions and vasospastic responses in pigs in vivo. The role of platelet-derived growth factor. *J Clin Invest*. 1996;97(3):769–76. <https://doi.org/10.1172/JCI118476>.
3. Sato M, Tani E, Fujikawa H, Kaibuchi K. Involvement of Rho-kinase-mediated phosphorylation of myosin light chain in enhancement of cerebral vasospasm. *Circ Res*. 2000;87(3):195–200. <https://www.ahajournals.org/doi/10.1161/01.RES.87.3.195>.
4. Takagi Y, Yasuda S, Takahashi J, Takeda M, Nakayama M, Ito K, Hirose M, Wakayama Y, Fukuda K, Shimokawa H. Importance of dual induction tests for coronary vasospasm and ventricular fibrillation in patients surviving out-of-hospital cardiac arrest. *Circ J*. 2009;73(4):767–9. <https://doi.org/10.1253/circj.CJ-09-0061>.
5. Hizume T, Morikawa K, Takaki A, Abe K, Sunagawa K, Amano M, Kaibuchi K, Kubo C, Shimokawa H. Sustained elevation of serum cortisol level causes sensitization of coronary vasoconstricting responses in pigs in vivo: a possible link between stress and coronary vasospasm. *Circ Res*. 2006;99(7):767–75. <https://doi.org/10.1161/01.RES.0000244093.69985.2f>.
6. Kandabashi T, Shimokawa H, Miyata K, Kunihiro I, Kawano Y, Fukata Y, Higo T, Egashira K, Takahashi S, Kaibuchi K, Takeshita A. Inhibition of myosin phosphatase by upregulated Rho-kinase plays a key role for coronary artery spasm in a porcine model with interleukin-1 $\beta$ . *Circulation*. 2000;101(11):1319–23. <https://doi.org/10.1161/01.CIR.101.11.1319>.
7. Katsumata N, Shimokawa H, Seto M, Kozai T, Yamawaki T, Kuwata K, Egashira K, Ikegaki I, Asano T, Sasaki Y, Takeshita A. Enhanced myosin light chain phosphorylations as a central mechanism for coronary artery spasm in a swine model with interleukin-1 $\beta$ . *Circulation*. 1997;96(12):4357–63. <https://doi.org/10.1161/01.cir.96.12.4357>.
8. Shimokawa H, Seto M, Katsumata N, Amano M, Kozai T, Yamawaki T, Kuwata K, Kandabashi T, Egashira K, Ikegaki I, Asano T, Kaibuchi K, Takeshita A. Rho-kinase-mediated pathway induces enhanced myosin light chain phosphorylations in a swine model of coronary artery spasm. *Cardiovasc Res*. 1999;43(4):1029–39. [https://doi.org/10.1016/s0008-6363\(99\)00144-3](https://doi.org/10.1016/s0008-6363(99)00144-3).
9. Masumoto A, Mohri M, Shimokawa H, Urakami L, Usui M, Takeshita A. Suppression of coronary artery spasm by the Rho-kinase inhibitor fasudil in patients with vasospastic angina. *Circulation*. 2002;105(13):1545–7. <https://doi.org/10.1161/hc1002.105938>.
10. Mohri M, Shimokawa H, Hiraoka Y, Masumoto A, Takeshita A. Rho-kinase inhibition with intracoronary fasudil prevents myocardial ischemia in patients with coronary microvascular spasm. *J Am Coll Cardiol*. 2003;41(1):15–9. [https://doi.org/10.1016/S0735-1097\(02\)02632-3](https://doi.org/10.1016/S0735-1097(02)02632-3).
11. Shimokawa H, Hiramori K, Inuma H, Hosoda S, Kishida H, Osada H, Katagiri T, Yamauchi K, Yui Y, Minamino T, Nakashima M, Kato K. Anti-anginal effect of fasudil, a Rho-kinase inhibitor, in patients with stable effort angina: a multicenter study. *J Cardiovasc Pharmacol*. 2002;40(5):751–61. <https://doi.org/10.1097/00005344-200211000-00013>.
12. Kikuchi Y, Yasuda S, Aizawa K, Tsuburaya R, Ito Y, Takeda M, Nakayama M, Ito K, Takahashi J, Shimokawa H. Enhanced Rho-kinase activity in circulating neutrophils of patients with vasospastic angina: a possible biomarker for diagnosis and disease activity assessment. *J Am Coll Cardiol*. 2011;58(12):1231–7. <https://doi.org/10.1016/j.jacc.2011.05.046>.
13. Fukata Y, Amano M, Kaibuchi K. Rho-Rho-kinase pathway in smooth muscle contraction and cytoskeletal reorganization of non-muscle cells. *Trends Pharmacol Sci*. 2001;22(1):32–9. [https://doi.org/10.1016/s0165-6147\(00\)01596-0](https://doi.org/10.1016/s0165-6147(00)01596-0).
14. Takai Y, Sasaki T, Matozaki T. Small GTP-binding proteins. *Physiol Rev*. 2001;81(1):153–208. <https://doi.org/10.1152/physrev.2001.81.1.153>.

15. Loirand G, Pacaud P. The role of Rho protein signaling in hypertension. *Nat Rev Cardiol.* 2010;7(11):637–47. <https://doi.org/10.1038/nrcardio.2010.136>.
16. Etienne-Manneville S, Hall A. Rho GTPases in cell biology. *Nature.* 2002;420(6916):629–35. <https://doi.org/10.1038/nature01148>.
17. Schmidt A, Hall A. Guanine nucleotide exchange factors for Rho GTPases: turning on the switch. *Genes Dev.* 2002;16(13):1587–609. <https://doi.org/10.1101/gad.1003302>.
18. Bernards A. GAPs galore! A survey of putative Ras superfamily GTPase activating proteins in man and Drosophila. *Biochim Biophys Acta.* 2003;1603(2):47–82. [https://doi.org/10.1016/s0304-419x\(02\)00082-3](https://doi.org/10.1016/s0304-419x(02)00082-3).
19. Olofsson B. Rho guanine dissociation inhibitors: pivotal molecules in cellular signalling. *Cell Signal.* 1999;11(8):545–54. [https://doi.org/10.1016/s0898-6568\(98\)00063-1](https://doi.org/10.1016/s0898-6568(98)00063-1).
20. Kimura K, Ito M, Amano M, Chihara K, Fukata Y, Nakafuku M, Yamamori B, Feng J, Nakano T, Okawa K, Iwamatsu A, Kaibuchi K. Regulation of myosin phosphatase by Rho and Rho-associated kinase (Rho-kinase). *Science.* 1996;273(5272):245–8. <https://doi.org/10.1126/science.273.5272.245>.
21. Ishizaki T, Maekawa M, Fujisawa K, Okawa K, Iwamatsu A, Fujita A, Watanabe N, Saito Y, Kakizuka A, Morii N, Narumiya S. The small GTP-binding protein Rho binds to and activates a 160 kDa Ser/Thr protein kinase homologous to myotonic dystrophy kinase. *EMBO J.* 1996;15(8):1885–93.
22. Leung T, Chen XQ, Manser E, Lim L. The p160 RhoA-binding kinase ROK $\alpha$  is a member of a kinase family and is involved in the reorganization of the cytoskeleton. *Mol Cell Biol.* 1996;16(10):5313–27. <https://doi.org/10.1128/mcb.16.10.5313>.
23. Shimokawa H, Sunamura S, Satoh K. RhoA/Rho-kinase in the cardiovascular system. *Circ Res.* 2016;118(2):352–66. <https://doi.org/10.1161/CIRCRESAHA.115.306532>.
24. Kina-Tanada M, Sakanashi M, Tanimoto A, Kaname T, Matsuzaki T, Noguchi K, Uchida T, Nakasone J, Kozuka C, Ishida M, Kubota H, Taira Y, Totsuka Y, Kina SI, Sunakawa H, Omura J, Satoh K, Shimokawa H, Yanagihara N, Maeda S, Ohya Y, Matsushita M, Masuzaki H, Arasaki A, Tsutsui M. Long-term dietary nitrite and nitrate deficiency causes the metabolic syndrome, endothelial dysfunction and cardiovascular death in mice. *Diabetologia.* 2017;60(6):1138–51. <https://doi.org/10.1007/s00125-017-4259-6>.
25. Shimokawa H. Primary endothelial dysfunction: atherosclerosis. *J Mol Cell Cardiol.* 1999;31(1):23–37. <https://doi.org/10.1006/jmcc.1998.0841>.
26. Vanhoutte PM. Endothelium-derived free radicals: for worse and for better. *J Clin Invest.* 2001;107(1):23–5. <https://doi.org/10.1172/JCI11832>.
27. Shimokawa H. 2014 Williams Harvey lecture: importance of coronary vasomotion abnormalities -from bench to bedside. *Eur Heart J.* 2014;35(45):3180–93. <https://doi.org/10.1093/eurheartj/ehu427>.
28. Shimokawa H, Satoh K. Light and dark of reactive oxygen species for vascular function. *J Cardiovasc Pharmacol.* 2015;65(5):412–8. <https://doi.org/10.1097/FJC.000000000000159>.
29. Shimokawa H, Satoh K. Vascular function. *Arterioscler Thromb Vasc Biol.* 2014;34(11):2359–62. <https://doi.org/10.1161/ATVBAHA.114.304119>.
30. Shimizu T, Fukumoto Y, Tanaka S, Satoh K, Ikeda S, Shimokawa H. Crucial role of ROCK2 in vascular smooth muscle cells for hypoxia-induced pulmonary hypertension in mice. *Arterioscler Thromb Vasc Biol.* 2013;33(12):2780–91. <https://doi.org/10.1161/ATVBAHA.113.301357>.
31. Do e Z, Fukumoto Y, Sugimura K, Miura Y, Tatebe S, Yamamoto S, Aoki T, Nochioka K, Nergui S, Yaoita N, Satoh K, Kondo M, Nakano M, Wakayama Y, Fukuda K, Nihei T, Kikuchi Y, Takahashi J, Shimokawa H. Rho-kinase activation in patients with heart failure. *Circ J.* 2013;77(10):2542–50. <https://doi.org/10.1253/circj.CJ-13-0397>.
32. Enkhjargal B, Godo S, Sawada A, Suvd N, Saito H, Noda K, Satoh K, Shimokawa H. Endothelial AMP-activated protein kinase regulates blood pressure and coronary flow responses through hyperpolarization mechanism in mice. *Arterioscler Thromb Vasc Biol.* 2014;34(7):1505–13. <https://doi.org/10.1161/ATVBAHA.114.303735>.

33. Shimokawa H, Takeshita A. Rho-kinase is an important therapeutic target in cardiovascular medicine. *Arterioscler Thromb Vasc Biol.* 2005;25(9):1767–75. <https://doi.org/10.1161/01.ATV.0000176193.83629.c8>.
34. Leung T, Manser E, Tan L, Lim L. A novel serine/threonine kinase binding the Ras-related RhoA GTPase which translocates the kinase to peripheral membranes. *J Biol Chem.* 1995;270(49):29051–4. <https://doi.org/10.1074/jbc.270.49.29051>.
35. Matsui T, Amano M, Yamamoto T, Chihara K, Nakafuku M, Ito M, Nakano T, Okawa K, Iwamatsu A, Kaibuchi K. Rho-associated kinase, a novel serine/threonine kinase, as a putative target for small GTP binding protein Rho. *EMBO J.* 1996;15(9):2208–16. <https://doi.org/10.1002/j.1460-2075.1996.tb00574.x>.
36. Nakagawa O, Fujisawa K, Ishizaki T, Saito Y, Nakao K, Narumiya S. ROCK-I and ROCK-II, two isoforms of Rho-associated coiled-coil forming protein serine/threonine kinase in mice. *FEBS Lett.* 1996;392(2):189–93. [https://doi.org/10.1016/0014-5793\(96\)00811-3](https://doi.org/10.1016/0014-5793(96)00811-3).
37. Uehata M, Ishizaki T, Satoh H, Ono T, Kawahara T, Morishita T, Tamakawa H, Yamagami K, Inui J, Maekawa M, Narumiya S. Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. *Nature.* 1997;389(6654):990–4. <https://doi.org/10.1038/40187>.
38. Davies SP, Reddy H, Caivano M, Cohen P. Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochem J.* 2000;351(Pt 1):95–105. <https://doi.org/10.1042/0264-6021:3510095>.
39. Higashi M, Shimokawa H, Hattori T, Hiroki J, Mukai Y, Morikawa K, Ichiki T, Takahashi S, Takeshita A. Long-term inhibition of Rho-kinase suppresses angiotensin II-induced cardiovascular hypertrophy in rats in vivo: effect on endothelial NAD(P)H oxidase system. *Circ Res.* 2003;93(8):767–75. <https://doi.org/10.1161/01.RES.0000096650.91688.28>.
40. Coleman ML, Sahai EA, Yeo M, Bosch M, Dewar A, Olson MF. Membrane blebbing during apoptosis results from caspase-mediated activation of ROCK I. *Nat Cell Biol.* 2001;3(4):339–45. <https://doi.org/10.1038/35070009>.
41. Sebbagh M, Hamelin J, Bertoglio J, Solary E, Breard J. Direct cleavage of ROCK II by granzyme B induces target cell membrane blebbing in a caspase-independent manner. *J Exp Med.* 2005;201(3):465–71. <https://doi.org/10.1084/jem.20031877>.
42. Komander D, Garg R, Wan PT, Ridley AJ, Barford D. Mechanism of multi-site phosphorylation from a ROCK-I:RhoE complex structure. *EMBO J.* 2008;27(23):3175–85. <https://doi.org/10.1038/emboj.2008.226>.
43. Wang Y, Zheng XR, Riddick N, Bryden M, Baur W, Zhang X, Surks HK. ROCK isoform regulation of myosin phosphatase and contractility in vascular smooth muscle cells. *Circ Res.* 2009;104(4):531–40. <https://doi.org/10.1161/CIRCRESAHA.108.188524>.
44. Riento K, Guasch RM, Garg R, Jin B, Ridley AJ. RhoE binds to ROCK I and inhibits downstream signaling. *Mol Cell Biol.* 2003;23(12):4219–29. <https://doi.org/10.1128/mcb.23.12.4219-4229.2003>.
45. Hiroki J, Shimokawa H, Higashi M, Morikawa K, Kandabashi T, Kawamura N, Kubota T, Ichiki T, Amano M, Kaibuchi K, Takeshita A. Inflammatory stimuli upregulate Rho-kinase in human coronary vascular smooth muscle cells. *J Mol Cell Cardiol.* 2004;37(2):537–46. <https://doi.org/10.1016/j.yjmcc.2004.05.008>.
46. Loirand G, Guerin P, Pacaud P. Rho kinases in cardiovascular physiology and pathophysiology. *Circ Res.* 2006;98(3):322–34. <https://doi.org/10.1161/01.RES.0000201960.04223.3c>.
47. Riento K, Ridley AJ. ROCKs: multifunctional kinases in cell behaviour. *Nat Rev Mol Cell Biol.* 2003;4(6):446–56. <https://doi.org/10.1038/nrm1128>.
48. Thumkeo D, Keel J, Ishizaki T, Hirose M, Nonomura K, Oshima H, Oshima M, Taketo MM, Narumiya S. Targeted disruption of the mouse Rho-associated kinase 2 gene results in intra-uterine growth retardation and fetal death. *Mol Cell Biol.* 2003;23(14):5043–55. <https://doi.org/10.1128/mcb.23.14.5043-5055.2003>.
49. Shimizu Y, Thumkeo D, Keel J, Ishizaki T, Oshima H, Oshima M, Noda Y, Matsumura F, Taketo MM, Narumiya S. ROCK-I regulates closure of the eyelids and ventral body wall

- by inducing assembly of actomyosin bundles. *J Cell Biol.* 2005;168(6):941–53. <https://doi.org/10.1083/jcb.200411179>.
50. Liao JK, Seto M, Noma K. Rho kinase (ROCK) inhibitors. *J Cardiovasc Pharmacol.* 2007;50(1):17–24. <https://doi.org/10.1097/FJC.0b013e318070d1bd>.
51. Noma K, Rikitake Y, Oyama N, Yan G, Alcaide P, Liu PY, Wang H, Ahl D, Sawada N, Okamoto R, Hiroi Y, Shimizu K, Lusinskas FW, Sun J, Liao JK. ROCK1 mediates leukocyte recruitment and neointima formation following vascular injury. *J Clin Invest.* 2008;118(5):1632–44. <https://doi.org/10.1172/JCI29226>.
52. Zhou Q, Gensch C, Liao JK. Rho-associated coiled-coil-forming kinases (ROCKs): potential targets for the treatment of atherosclerosis and vascular disease. *Trends Pharmacol Sci.* 2011;32(3):167–73. <https://doi.org/10.1016/j.tips.2010.12.006>.
53. Radeff JM, Nagy Z, Stern PH. Rho and Rho kinase are involved in parathyroid hormone-stimulated protein kinase C  $\alpha$  translocation and IL-6 promoter activity in osteoblastic cells. *J Bone Miner Res.* 2004;19(11):1882–91. <https://doi.org/10.1359/jbmr.040806>.
54. Funakoshi Y, Ichiki T, Shimokawa H, Egashira K, Takeda K, Kaibuchi K, Takeya M, Yoshimura T, Takeshita A. Rho-kinase mediates angiotensin II-induced monocyte chemoattractant protein-1 expression in rat vascular smooth muscle cells. *Hypertension.* 2001;38(1):100–4. <https://doi.org/10.1161/01.hyp.38.1.100>.
55. Hattori T, Shimokawa H, Higashi M, Hiroki J, Mukai Y, Kaibuchi K, Takeshita A. Long-term treatment with a specific Rho-kinase inhibitor suppresses cardiac allograft vasculopathy in mice. *Circ Res.* 2004;94(1):46–52. <https://doi.org/10.1161/01.RES.0000107196.21335.2B>.
56. Hattori T, Shimokawa H, Higashi M, Hiroki J, Mukai Y, Tsutsui H, Kaibuchi K, Takeshita A. Long-term inhibition of Rho-kinase suppresses left ventricular remodeling after myocardial infarction in mice. *Circulation.* 2004;109(18):2234–9. <https://doi.org/10.1161/01.CIR.0000127939.16111.58>.
57. Wang F, Okamoto Y, Inoki I, Yoshioka K, Du W, Qi X, Takuwa N, Gonda K, Yamamoto Y, Ohkawa R, Nishiuchi T, Sugimoto N, Yatomi Y, Mitsumori K, Asano M, Kinoshita M, Takuwa Y. Sphingosine-1-phosphate receptor-2 deficiency leads to inhibition of macrophage proinflammatory activities and atherosclerosis in apoE-deficient mice. *J Clin Invest.* 2010;120(11):3979–95. <https://doi.org/10.1172/jci42315>.
58. Takemoto M, Sun J, Hiroki J, Shimokawa H, Liao JK. Rho-kinase mediates hypoxia-induced downregulation of endothelial nitric oxide synthase. *Circulation.* 2002;106(1):57–62. <https://doi.org/10.1161/01.CIR.0000020682.73694.AB>.
59. Oi K, Shimokawa H, Hiroki J, Uwatoku T, Abe K, Matsumoto Y, Nakajima Y, Nakajima K, Takeichi S, Takeshita A. Remnant lipoproteins from patients with sudden cardiac death enhance coronary vasospastic activity through upregulation of Rho-kinase. *Arterioscler Thromb Vasc Biol.* 2004;24(5):918–22. <https://doi.org/10.1161/01.atv.0000126678.93747.80>.
60. Mackay DJ, Hall A. Rho GTPases. *J Biol Chem.* 1998;273(33):20685–8. <https://doi.org/10.1074/jbc.273.33.20685>.
61. Neco P, Giner D, Viniegra S, Borges R, Villarroel A, Gutierrez LM. New roles of myosin II during vesicle transport and fusion in chromaffin cells. *J Biol Chem.* 2004;279(26):27450–7. <https://doi.org/10.1074/jbc.M311462200>.
62. Omar HA, Cherry PD, Mortelliti MP, Burke-Wolin T, Wolin MS. Inhibition of coronary artery superoxide dismutase attenuates endothelium-dependent and -independent nitrovasodilator relaxation. *Circ Res.* 1991;69(3):601–8. <https://doi.org/10.1161/01.RES.69.3.601>.
63. Alexander RW. Theodore Cooper memorial lecture. Hypertension and the pathogenesis of atherosclerosis. Oxidative stress and the mediation of arterial inflammatory response: a new perspective. *Hypertension.* 1995;25(2):155–61. <https://doi.org/10.1161/01.hyp.25.2.155>.
64. Baas AS, Berk BC. Differential activation of mitogen-activated protein kinases by H<sub>2</sub>O<sub>2</sub> and O<sub>2</sub><sup>-</sup> in vascular smooth muscle cells. *Circ Res.* 1995;77(1):29–36. <https://doi.org/10.1161/01.res.77.1.29>.
65. Griendling KK, Minieri CA, Ollerenshaw JD, Alexander RW. Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ Res.* 1994;74(6):1141–8. <https://doi.org/10.1161/01.res.74.6.1141>.



66. Takeda K, Ichiki T, Tokunou T, Iino N, Fujii S, Kitabatake A, Shimokawa H, Takeshita A. Critical role of Rho-kinase and MEK/ERK pathways for angiotensin II-induced plasminogen activator inhibitor type-1 gene expression. *Arterioscler Thromb Vasc Biol.* 2001;21(5):868–73. <https://doi.org/10.1161/01.ATV.21.5.868>.
67. Guilluy C, Bregeon J, Toumaniantz G, Rolli-Derkinderen M, Retailliau K, Loufrani L, Henrion D, Scalbert E, Bril A, Torres RM, Offermanns S, Pacaud P, Loirand G. The Rho exchange factor Arhgef1 mediates the effects of angiotensin II on vascular tone and blood pressure. *Nat Med.* 2010;16(2):183–90. <https://doi.org/10.1038/nm.2079>.
68. Seasholtz TM, Majumdar M, Kaplan DD, Brown JH. Rho and Rho kinase mediate thrombin-stimulated vascular smooth muscle cell DNA synthesis and migration. *Circ Res.* 1999;84(10):1186–93. <https://doi.org/10.1161/01.RES.84.10.1186>.
69. van Nieuw Amerongen GP, van Delft S, Vermeer MA, Collard JG, van Hinsbergh VW. Activation of RhoA by thrombin in endothelial hyperpermeability: role of Rho-kinase and protein tyrosine kinases. *Circ Res.* 2000;87(4):335–40. <https://doi.org/10.1161/01.RES.87.4.335>.
70. Kishi H, Bao J, Kohama K. Inhibitory effects of ML-9, wortmannin, and Y-27632 on the chemotaxis of vascular smooth muscle cells in response to platelet-derived growth factor-BB. *J Biochem.* 2000;128(5):719–22. <https://doi.org/10.1093/oxfordjournals.jbchem.a022806>.
71. Sauzeau V, Le Jeune H, Cario-Toumaniantz C, Vaillant N, Gadeau AP, Desgranges C, Scalbert E, Chardin P, Pacaud P, Loirand G. P2Y<sub>1</sub>, P2Y<sub>2</sub>, P2Y<sub>4</sub>, and P2Y<sub>6</sub> receptors are coupled to Rho and Rho kinase activation in vascular myocytes. *Am J Physiol Heart Circ Physiol.* 2000;278(6):H1751–61. <https://doi.org/10.1152/ajpheart.2000.278.6.H1751>.
72. Sauzeau V, Le Mellionnec E, Bertoglio J, Scalbert E, Pacaud P, Loirand G. Human urotensin II-induced contraction and arterial smooth muscle cell proliferation are mediated by RhoA and Rho-kinase. *Circ Res.* 2001;88(11):1102–4. <https://doi.org/10.1161/hh1101.092034>.
73. Takemoto M, Liao JK. Pleiotropic effects of 3-hydroxy-3-methylglutaryl coenzyme a reductase inhibitors. *Arterioscler Thromb Vasc Biol.* 2001;21(11):1712–9. <https://doi.org/10.1161/hq1101.098486>.
74. Shimokawa H. Cellular and molecular mechanisms of coronary artery spasm: lessons from animal models. *Jpn Circ J.* 2000;64(1):1–12. [https://doi.org/10.1007/978-3-642-56225-9\\_56](https://doi.org/10.1007/978-3-642-56225-9_56).
75. Shimokawa H. Rho-kinase as a novel therapeutic target in treatment of cardiovascular diseases. *J Cardiovasc Pharmacol.* 2002;39(3):319–27. <https://doi.org/10.1097/00005344-200203000-00001>.
76. Amano M, Chihara K, Kimura K, Fukata Y, Nakamura N, Matsuura Y, Kaibuchi K. Formation of actin stress fibers and focal adhesions enhanced by Rho-kinase. *Science.* 1997;275(5304):1308–11. <https://doi.org/10.1126/science.275.5304.1308>.
77. Hall A. Rho GTPases and the actin cytoskeleton. *Science.* 1998;279(5350):509–14. <https://doi.org/10.1126/science.279.5350.509>.
78. Rao GN, Berk BC. Active oxygen species stimulate vascular smooth muscle cell growth and proto-oncogene expression. *Circ Res.* 1992;70(3):593–9. <https://doi.org/10.1161/01.res.70.3.593>.
79. Ross R. Atherosclerosis is an inflammatory disease. *Am Heart J.* 1999;138(5 Pt 2):S419–20. [https://doi.org/10.1016/s0002-8703\(99\)70266-8](https://doi.org/10.1016/s0002-8703(99)70266-8).
80. Libby P. Inflammation in atherosclerosis. *Nature.* 2002;420(6917):868–74. <https://doi.org/10.1038/nature01323>.
81. Inoue T, Node K. Molecular basis of restenosis and novel issues of drug-eluting stents. *Circ J.* 2009;73(4):615–21. <https://doi.org/10.1253/circj.cj-09-0059>.
82. Shimokawa H, Rashid M. Development of Rho-kinase inhibitors for cardiovascular medicine. *Trends Pharmacol Sci.* 2007;28(6):296–302. <https://doi.org/10.1016/j.tips.2007.04.006>.
83. Eto Y, Shimokawa H, Hiroki J, Morishige K, Kandabashi T, Matsumoto Y, Amano M, Hoshijima M, Kaibuchi K, Takeshita A. Gene transfer of dominant negative Rho-kinase suppresses neointimal formation after balloon injury in pigs. *Am J Physiol Heart Circ Physiol.* 2000;278(6):H1744–50. <https://doi.org/10.1152/ajpheart.2000.278.6.H1744>.



84. Sawada N, Itoh H, Ueyama K, Yamashita J, Doi K, Chun TH, Inoue M, Masatsugu K, Saito T, Fukunaga Y, Sakaguchi S, Arai H, Ohno N, Komeda M, Nakao K. Inhibition of Rho-associated kinase results in suppression of neointimal formation of balloon-injured arteries. *Circulation*. 2000;101(17):2030–3. <https://doi.org/10.1161/01.cir.101.17.2030>.
85. Shibata R, Kai H, Seki Y, Kato S, Morimatsu M, Kaibuchi K, Imaizumi T. Role of Rho-associated kinase in neointima formation after vascular injury. *Circulation*. 2001;103(2):284–9. <https://doi.org/10.1161/01.CIR.103.2.284>.
86. Miyata K, Shimokawa H, Kandabashi T, Higo T, Morishige K, Eto Y, Egashira K, Kaibuchi K, Takeshita A. Rho-kinase is involved in macrophage-mediated formation of coronary vascular lesions in pigs in vivo. *Arterioscler Thromb Vasc Biol*. 2000;20(11):2351–8. <https://doi.org/10.1161/01.atv.20.11.2351>.
87. Shimokawa H, Morishige K, Miyata K, Kandabashi T, Eto Y, Ikegaki I, Asano T, Kaibuchi K, Takeshita A. Long-term inhibition of Rho-kinase induces a regression of arteriosclerotic coronary lesions in a porcine model in vivo. *Cardiovasc Res*. 2001;51(1):169–77. [https://doi.org/10.1016/S0008-6363\(01\)00291-7](https://doi.org/10.1016/S0008-6363(01)00291-7).
88. Matsumoto Y, Uwatoku T, Oi K, Abe K, Hattori T, Morishige K, Eto Y, Fukumoto Y, Nakamura K, Shibata Y, Matsuda T, Takeshita A, Shimokawa H. Long-term inhibition of Rho-kinase suppresses neointimal formation after stent implantation in porcine coronary arteries: involvement of multiple mechanisms. *Arterioscler Thromb Vasc Biol*. 2004;24(1):181–6. <https://doi.org/10.1161/01.atv.0000105053.46994.5b>.
89. Kandabashi T, Shimokawa H, Miyata K, Kunihiro I, Eto Y, Morishige K, Matsumoto Y, Obara K, Nakayama K, Takahashi S, Takeshita A. Evidence for protein kinase C-mediated activation of Rho-kinase in a porcine model of coronary artery spasm. *Arterioscler Thromb Vasc Biol*. 2003;23(12):2209–14. <https://doi.org/10.1161/01.ATV.0000104010.87348.26>.
90. Yamakawa T, Tanaka S, Numaguchi K, Yamakawa Y, Motley ED, Ichihara S, Inagami T. Involvement of Rho-kinase in angiotensin II-induced hypertrophy of rat vascular smooth muscle cells. *Hypertension*. 2000;35(1 Pt 2):313–8. <https://doi.org/10.1161/01.HYP.35.1.313>.
91. Kandabashi T, Shimokawa H, Mukai Y, Matoba T, Kunihiro I, Morikawa K, Ito M, Takahashi S, Kaibuchi K, Takeshita A. Involvement of Rho-kinase in agonists-induced contractions of arteriosclerotic human arteries. *Arterioscler Thromb Vasc Biol*. 2002;22(2):243–8. <https://doi.org/10.1161/hq0202.104274>.
92. Yada T, Shimokawa H, Hiramatsu O, Kajita T, Shigeto F, Tanaka E, Shinozaki Y, Mori H, Kiyooka T, Katsura M, Ohkuma S, Goto M, Ogasawara Y, Kajiya F. Beneficial effect of hydroxyfasudil, a specific Rho-kinase inhibitor, on ischemia/reperfusion injury in canine coronary microcirculation in vivo. *J Am Coll Cardiol*. 2005;45(4):599–607. <https://doi.org/10.1016/j.jacc.2004.10.053>.
93. Sato S, Ikegaki I, Asano T, Shimokawa H. Antiischemic properties of fasudil in experimental models of vasospastic angina. *Jpn J Pharmacol*. 2001;87(1):34–40. <https://doi.org/10.1254/jjp.87.34>.
94. Utsunomiya T, Satoh S, Ikegaki I, Toshima Y, Asano T, Shimokawa H. Antianginal effects of hydroxyfasudil, a Rho-kinase inhibitor, in a canine model of effort angina. *Br J Pharmacol*. 2001;134(8):1724–30. <https://doi.org/10.1038/sj.bjp.0704410>.
95. Satoh S, Ikegaki I, Toshima Y, Watanabe A, Asano T, Shimokawa H. Effects of Rho-kinase inhibitor on vasopressin-induced chronic myocardial damage in rats. *Life Sci*. 2002;72(1):103–12. [https://doi.org/10.1016/s0024-3205\(02\)02178-1](https://doi.org/10.1016/s0024-3205(02)02178-1).
96. Mehta PK, Griendling KK. Angiotensin II cell signaling: physiological and pathological effects in the cardiovascular system. *Am J Physiol Cell Physiol*. 2007;292(1):C82–97. <https://doi.org/10.1152/ajpcell.00287.2006>.
97. Sadoshima J, Xu Y, Slayter HS, Izumo S. Autocrine release of angiotensin II mediates stretch-induced hypertrophy of cardiac myocytes in vitro. *Cell*. 1993;75(5):977–84. [https://doi.org/10.1016/0092-8674\(93\)90541-w](https://doi.org/10.1016/0092-8674(93)90541-w).
98. Nakamura K, Fushimi K, Kouchi H, Mihara K, Miyazaki M, Ohe T, Namba M. Inhibitory effects of antioxidants on neonatal rat cardiac myocyte hypertrophy induced by tumor necrosis factor- $\alpha$  and angiotensin II. *Circulation*. 1998;98(8):794–9. <https://doi.org/10.1161/01.CIR.98.8.794>.

99. Akki A, Zhang M, Murdoch C, Brewer A, Shah AM. NADPH oxidase signaling and cardiac myocyte function. *J Mol Cell Cardiol.* 2009;47(1):15–22. <https://doi.org/10.1016/j.yjmcc.2009.04.004>.
100. Shibata R, Ouchi N, Murohara T. Adiponectin and cardiovascular disease. *Circ J.* 2009;73(4):608–14. <https://doi.org/10.1253/circj.CJ-09-0057>.
101. Vahebi S, Kobayashi T, Warren CM, de Tombe PP, Solaro RJ. Functional effects of Rho-kinase-dependent phosphorylation of specific sites on cardiac troponin. *Circ Res.* 2005;96(7):740–7. <https://doi.org/10.1161/01.RES.0000162457.56568.7d>.
102. Fukui S, Fukumoto Y, Suzuki J, Saji K, Nawata J, Tawara S, Shinozaki T, Kagaya Y, Shimokawa H. Long-term inhibition of Rho-kinase ameliorates diastolic heart failure in hypertensive rats. *J Cardiovasc Pharmacol.* 2008;51(3):317–26. <https://doi.org/10.1097/FJC.0b013e31816533b7>.
103. Kishi T, Hirooka Y, Masumoto A, Ito K, Kimura Y, Inokuchi K, Tagawa T, Shimokawa H, Takeshita A, Sunagawa K. Rho-kinase inhibitor improves increased vascular resistance and impaired vasodilation of the forearm in patients with heart failure. *Circulation.* 2005;111(21):2741–7. <https://doi.org/10.1161/CIRCULATIONAHA.104.510248>.
104. Mukai Y, Shimokawa H, Matoba T, Kandabashi T, Satoh S, Hiroki J, Kaibuchi K, Takeshita A. Involvement of Rho-kinase in hypertensive vascular disease: a novel therapeutic target in hypertension. *FASEB J.* 2001;15(6):1062–4. <https://doi.org/10.1096/fsb2fj000735fje>.
105. Ito K, Hirooka Y, Sakai K, Kishi T, Kaibuchi K, Shimokawa H, Takeshita A. Rho/Rho-kinase pathway in brain stem contributes to blood pressure regulation via sympathetic nervous system: possible involvement in neural mechanisms of hypertension. *Circ Res.* 2003;92(12):1337–43. <https://doi.org/10.1161/01.RES.0000079941.59846.D4>.
106. Ito K, Hirooka Y, Kishi T, Kimura Y, Kaibuchi K, Shimokawa H, Takeshita A. Rho/Rho-kinase pathway in the brainstem contributes to hypertension caused by chronic nitric oxide synthase inhibition. *Hypertension.* 2004;43(2):156–62. <https://doi.org/10.1161/01.HYP.0000114602.82140.a4>.
107. Abe K, Shimokawa H, Morikawa K, Uwatoku T, Oi K, Matsumoto Y, Hattori T, Nakashima Y, Kaibuchi K, Sueishi K, Takeshita A. Long-term treatment with a Rho-kinase inhibitor improves monocrotaline-induced fatal pulmonary hypertension in rats. *Circ Res.* 2004;94(3):385–93. <https://doi.org/10.1161/01.RES.0000111804.34509.94>.
108. Abe K, Tawara S, Oi K, Hizume T, Uwatoku T, Fukumoto Y, Kaibuchi K, Shimokawa H. Long-term inhibition of Rho-kinase ameliorates hypoxia-induced pulmonary hypertension in mice. *J Cardiovasc Pharmacol.* 2006;48(6):280–5. <https://doi.org/10.1097/01.fjc.0000248244.64430.4a>.
109. Do e Z, Fukumoto Y, Takaki A, Tawara S, Ohashi J, Nakano M, Tada T, Saji K, Sugimura K, Fujita H, Hoshikawa Y, Nawata J, Kondo T, Shimokawa H. Evidence for Rho-kinase activation in patients with pulmonary arterial hypertension. *Circ J.* 2009;73(9):1731–9. <https://doi.org/10.1253/circj.cj-09-01350>.
110. Fukumoto Y, Matoba T, Ito A, Tanaka H, Kishi T, Hayashidani S, Abe K, Takeshita A, Shimokawa H. Acute vasodilator effects of a Rho-kinase inhibitor, fasudil, in patients with severe pulmonary hypertension. *Heart.* 2005;91(3):391–2. <https://doi.org/10.1007/s00380-009-1176-8>.